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Neurological Diseases Associated with Brain Iron Accumulation

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Abstract

Brain iron plays a very important role in maintaining normal physiological functioning and homeostasis. However, excess iron or dysregulated iron metabolism is a potent source of free radical formation and oxidative damage to neuronal and other brain cells. Abnormal high brain iron levels are associated with many neurological diseases like Alzheimer's disease, Parkinson's disease, Huntington's disease, Neurodegeneration with brain iron accumulation, Multiple sclerosis, Aceruloplasminaemia (CPM) etc. Here in this review we will focus on brain iron transport, dysregulated metabolism and the disease it causes.

Keywords: Brain Iron; Neurodegeneration; Neurodegenerative Mechanisms; Neuroinflammation; Mitochondria; Neuron; Cells; Iron Metabolism; Blood Brain Barrier; Blood CSF Barrier; Influx; Efflux

Abbreviation

DMT: Divalent Metal (Ion) Transporter; FPN: Ferroportin; CP: Ceruloplasmin; Heph: Hephaestin; SDR2: Stromal Cell-Derived Receptor; Nfts: Neurofibrillary Tangles; APP: Amyloid Precursor Protein; IRP-IRE: Iron Regulatory Protein-Iron Responsive Element; Fe: Iron; PANK: Pantothenate Kinase; FTL: Ferritin Light Chain; ROS: Reactive Oxygen Species; FRDA: Friedreich's Ataxia; FXN: Frataxin; ICH: Intracerebral Hemorrhage; GPX: Glutathione Peroxidase

Introduction

Iron is the most abundant metal in the brain [1,2]. It is responsible for many cellular activities like mitochondrial respiration, synthesis of myelin and even synthesis of neurotransmitter and its metabolism. Iron in brain plays a crucial physiological role in maintaining homeostasis [3,4]. Brain iron status is consistently maintained and tightly regulated at the level of the blood brain barrier (BBB)[5] and blood cerebrospinal fluid barrier(BCSFB) [6,7]. So it is very important for us to understand the impact of iron deficiency in development of brain and excess causing diseases which are mostly degenerative in nature [8]. May be detailed understanding the role of BBB and mechanism of transport of iron across it is vital.

The normal structure and function of the BBB is essential for brain iron homeostasis because the endothelial cells of the BBB are a regulatory site for brain iron uptake [9]. BBB endothelial cells can also regulate the two possible iron transport pathways[transferrinbound iron (Tf-Fe) and non-transferrin-bound iron (NTBI)] by controlling receptor expression, internalization of transferrin receptor 1(TfR1) complexes, and acidification inside cell endosomes [10]. The mechanism involves two transmembrane steps: iron uptake into the microvascular endothelial cells at the luminal membrane (apical, on the blood side), followed by iron efflux into the brain interstitium at the abluminal membrane (basal, brain side) [6]. The accumulated evidence suggests that the Tf/TfR1 pathway is the major route or primary pathway for iron transport across the luminal membrane of the capillary endothelium [11] and that iron, possibly in the form of Fe2+, crosses the abluminal membrane and enters the brain parenchyma [12,13].

Apart from this two pathway there may be some other ways of transportation of iron in brain a)H-ferritin-mediated delivery of iron into the brain [14] b) transcytosis of Tf/TfR1 complexes through brain endothelial cells, leading to the release of iron from the abluminal side into the brain [15] c) lactoferrin-lactoferrin recep-

tor (Lf/LfR) [16] d) secreted melanotransferrin (p97)-glycosylphosphatidylinositol (GPI)-anchored p97(sP97/GPI-P97) [17].

Blood-Cerebrospinal Fluid Barrier (BCSFB) also separates systemic circulation from the brain parenchyma physically [6]. Tf/TfR1/DMT1 pathway may be of importance for iron transport across the BCSFB, and also that the export of iron from the choroid epithelium to the cerebrospinal fluid (CSF) is mediated by the Fpn1/CP or Fpn1/Heph pathways [7]. Once iron enters the interstitial fluid or ventricular CSF, it binds to apo-Tf, synthesized locally by the choroid plexus [11]. SDR2 is the only known ferric reductase expressed in the choroid plexus and that DMT1 is also present in the choroid plexus it is possible that iron reduction by this protein followed by DMT1-mediated absorption might be an alternative mechanism for iron transport across the BCSFB [18,19]. It is possible that the BCSFB is used more for iron removal from the brain than iron transport into the brain [20]. There is a diurnal variation of iron content in the ventral midbrain supporting the existence of mechanisms for iron export from the brain to the systemic compartment [21]. It has been suggested that the glymphatic system in the brain [22,23] may be one of the routes by which transition metals including iron are transported and cleared from the brain via convective bulk flow of interstitial fluid, possibly mediated by astrocytic end-feet-expressed aquaporin-4 (Aqp4) water channels [2]. Iron transport into the brain bypassing the brain barriers has also been suggested [24]. The circumventricular organs that receive a plethora of neuronal projections, mainly from hypothalamic nuclei, have been suggested to play a role in iron transport in the developing brain. Some motor neurons that project to peripheral organs devoid of a blood barrier express TfR1 and show retrograde axonal transport of iron into the brain [25].

Iron transport within the brain comprises of a) Tf-Fe and NTBI transport forms [11,24,26] b)uptake by neurons [27,28] c)uptake by oligodendrocytes [12] d) uptake by astrocytes [29] e) uptake by microglia [11] f) export from neurons [30], astrocytes [31], oligodendrocytes and microglia [32]. Fpn1 is the only known cellular iron expressed in neurons, astrocytes, oligodendrocytes, and microglia [32]. Transcriptional changes in Fpn1 mRNA were found to be mediated by hepcidin in differentiated neuronal-like PC12 cells subjected to iron challenge [33] and iron export through Fpn1 is modulated by the iron chaperone poly(rC)-binding protein 2 (PCBP2) [34]. CP, a critical ferroxidase, has been found in neurons and astrocytes [35], while another ferroxidase Heph is expressed in neurons [30], oligodendrocytes and microglia [32]. Iron efflux from the neuron is mediated by the Fpn1/CP and/or Fpn1/Heph pathways, from astrocytes by Fpn1/CP, and from oligodendrocytes and microglia by the Fpn1/Heph route.

Diseased caused by dysregulated iron metabolism Alzheimer's disease (AD)

Elevated iron in AD brains was first reported in 1953 [36] and shown to be associated with senile plaques(SPs). Iron is enriched in both NFTs [37] and SPs [36]. Iron accumulation occurs in AD cortex, but not cerebellum. The iron storage protein ferritin binds most iron within the brain [38], and this protein increases with age and in AD [39]. APP-knock out mice exhibit iron accumulation in brain and peripheral tissues, and loss of APP ferroxidase activity in AD brain is coincident with iron retention in the tissue. Iron-export capability of APP requires tau [40].Tau has been implicated in axonal transport [41] and binds APP [42]. Loss of tau in mice causes age-dependent iron accumulation [40]. Genetic factors could also increase the susceptibility to iron burden in AD like Tf variant C2, Mutations in the hemochromatosis gene(H63D and C82Y).

Parkinson's disease(PD)

Several studies reported that iron deposition was increased in the substantia nigra (SN) according to the severity of the disease in PD patients [43]. In PD brain, histology studies showed that iron accumulate in neurons and glia in SN [44]. Furthermore, there are reports that a dysfunction in the IRP-IRE system that results in iron accumulation gave rise to alpha synuclein(a-Syn) induced toxicity [45,46], that led to pathogenesis in PD [47]. Similarly, in almost all PD patient brains the Lewy bodies contained aggregated a-Syn [48]. Cellular iron accumulation in PD brain may be caused by elevated influx or decreased efflux. Inflammation could contribute to iron accumulation by either increasing DMT1 uptake activity or TfR transport activity. In a mouse model, DMT1 activity was increased to mediate the iron uptake [49], and this increase may be due to direct S-nitrosylation of DMT1 [50]. In addition, the presence of LfR on neurons and the increased expression of LfR on iron-induced degenerative dopaminergic neurons in PD which may imply that an Lf/LfR-mediated pathway also may be involved iniron (Lf-Fe) uptake by neurons [51]. Ferritin can hold large amount of iron as compared to Tf and if this physiological pathway gets supersaturated with iron can lead to accumulation and nigral pathology [52,53]. The reduced expression of Fpn1 has also been connected with the activation of N-methyl-D-aspartate (NMDA) receptor (NR)-induced iron accumulation and neurodegeneration in dopamine (DA) neurons in Parkinson's disease [54].

Huntington's disease(HD)

Impairment in intracellular iron levels and energy metabolism are both features of HD pathogenesis as HTT has been reported to mediate endocytosed Fe(II) required for oxidative energy production [55,56]. Magnetic resonance imaging(MRI) studies suggest

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changes in the metabolism of brain iron during early HD [57,58]. Post-mortem studies revealed increased levels of Fe(II) in the brains of HD individuals as compared to age-matched controls [59].

Neurodegeneration with brain iron accumulation (NBIA)

Neurodegeneration with brain iron accumulation comprises a number of pediatric and adult neurological diseases characterized by parkinsonism, dementia, and excessive iron accumulation in the basal ganglia [60]. Distinct genetic causes have been described within this disease spectrum yet no direct link between neurodegeneration and iron accumulation is proven. Mutations in PANK2, a gene implicated in coenzyme A biosynthesis, synthesis of lipids and citric acid cycle [61], results in panthoten kinase-associated neurodegeneration (PKAN). Though this mutation may not affect iron homeostasis directly, subsequent iron accumulation suggests that mitochondrial dysfunction may trigger iron dyshomeostasis. Although NBIA disorders are clinically characterized by the hallmark deposition of iron in the brain, for the majority of these disorders, this feature is a secondary consequence of the primary disease mechanism [62]. Detection of de novo mutations in WDR45—which encodes WD repeat domain 45, a β -propeller scaffolding protein with a presumed function in autophagy-in patients with β - propeller protein-associated neurodegeneration (BPAN) provides evidence of the association of defective autophagy with NBIA [63,64].

Multiple sclerosis (MS)

Iron content is elevated in deep grey matter structures and in the vicinity of lesions whereas it is reduced in the normal-appearing white matter (NAWM), and the extent of iron depletion correlates with disease duration. Furthermore, iron content is low in remyelinated plaques, suggesting that dynamic shuttling of iron continues throughout the MS disease process [65]. Elevated iron content has been detected using susceptibility MRI in deep grey matter structures, and is associated with increased disability and grey matter atrophy. In support of these MRI results, neuropathology of brain autopsy revealed substantial degeneration of deep grey matter structures, which corresponds with iron accumulation and oxidative damage [66]. Comparing measures from the same individual before and after death, changes indicative of iron accumulation detected by susceptibility MRI have been shown to correlate with increased iron content revealed by postmortem histopathology [67] or X-ray fluorescence [68]. In the vicinity of lesions, susceptibility MRI has revealed in vivo changes that are suggestive of iron deposition in areas that demonstrate myelin loss, focal iron

deposits or both at autopsy [69,70]. Patients with MS show evidence of widespread mitochondrial damage [71,72], and the consequent increase in production of reactive oxygen species can promote iron-mediated oxidative damage that leads to progressive genomic damage and further impairment of mitochondrial function [73]. Oligodendrocyte progenitor cells (OPCs), which are implicated in remyelination in MS, have heightened susceptibility to metabolic stress compared with oligodendrocytes [74]. In MS, iron accumulates in macrophages and microglia around the rim of lesions, and these iron-laden cells show signs of dystrophy [75].

Aceruloplasminemia

Aceruloplasminemia is caused by mutations in CP, encoding ceruloplasmin [76], an enzyme that catalyzes the peroxidation of ferrous transferrin to ferric transferrin. Aceruloplasminemia presents in the third to fifth decade of life with a progressive motor disorder, dementia, diabetes mellitus, and retinal degeneration, with evidence of brain iron deposition in the basal ganglia [77]. Some of the mutant CP cell proteins that exhibit impaired copper incorporation are able to transform into a CP holoprotein in the presence of copper-glutathione, suggesting a disruption of the intracellular copper-loading process [78]. The culmination of these abnormal cellular processes is the inability to prevent internalization and degradation of ferroprotein, leading to defective iron efflux and increased intracellular iron [78-80].

Neuroferritinopathy

Neuroferritinopathy may present from adolescence to the sixth decade, commonly with a progressive extrapyramidal disorder with radiological features of iron deposition in the basal ganglia and cystic degeneration. FTL is one of the two subunits of the main intracellular iron storage protein ferritin, and mutations in the encoding gene FTL cause neuroferritinopathy [81]. Pathological analyses reveal abnormal nuclear and cytoplasmic ferritin inclusion bodies in glia and neurons of the CNS as well as in other organs in association with iron accumulation [82,83]. Furthermore, studies have found an increase in ubiquitinated proteins and redistribution of proteasome components to the site of ferritin inclusions [83]. Mouse model also showed altered gene expression profiles for proteins involved in iron homeostasis, including decreased TfR-1 (the mouse ortholog of human TFRC) and Irp1 (the mouse ortholog of human IRP1) as well as markers of oxidative stress, such as lipid peroxidation products, oxidatively modified proteins, and protein radicals [82]. Although abnormal FTL is able to coassemble with endogenous ferritin heavy and light chains, the resultant ferritin molecules exhibit reduced efficiency for iron sequestration. This

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impairment in ferritin function results in an increase in the cellular labile iron pool along with enhanced ROS production, increased oxidized protein levels, and decreased proteasomal activity [84]. Defective FTL function affects normal iron metabolism, with secondary oxidative cell damage implicated as a mechanism in neurodegeneration.

Friedreich's ataxia

Iron relation was first evidenced for myocardial iron deposits in FRDA hearts [85]. Much later, iron involvement was recognized by the finding of deficiencies of aconitase and succinate dehydrogenase [86], both of which are mitochondrial iron–sulfur cluster-containing enzymes. FXN deficiency leads to mitochondrial iron overload, defective energy supply, and generation of reactive oxygen species [87].

Stroke

Growing evidence indicates that iron accumulation in neurons following ischemia increases brain susceptibility to iron-induced damage [88-90]. Red blood cell release and lysis have been considered the major factors mediating iron-induced brain damage after ICH [91]. Collagenase, which is commonly used to induce ICH, results in iron overload [92]. Moreover, iron overload may contribute to brain edema following ICH [93].

Amyotrophic lateral sclerosis (ALS)

A recent study firstly showed that the serum iron, ferritin, and transferrin saturation coefficient were significantly elevated in ALS patients, and the iron status was associated with body weight loss [94]. Serum ferritin may be a candidate biomarker for ALS aggravation [95]. Free iron level was also increased in the cerebrospinal fluid of ALS patients compared to controls [96,97]. A two fold increased level of inappropriate iron ligands in cerebrospinal fluid (CSF) was also found in patients with ALS, which may increase iron redox activity and reactive oxygen species production [98]. Autopsy study demonstrated that iron load in gray matter from the frontal cortex of ALS was increased significantly than that of controls [99]. The concentration of iron was also increased in the spinal cord of ALS patients [100-102]. Disruption in the expression of brain iron transporters such as lactotransferrin receptor, melanotransferrin, and ceruloplasmin is related to iron accumulation in ALS [103]. The deleterious effects on neuronal health and survival upon ablation of GPX4 in motor neurons might confer a role of ferroptosis to degenerative motor neuron diseases specially amyotrophic lateral sclerosis [103].

Sanfilippo syndrome

The reduced expression of Fpn1 has also been connected with the activation of N-methyl-D-aspartate (NMDA) receptor (NR)-induced iron accumulation and brain iron retention in Sanfilippo syndrome, a pediatric neurodegenerative disease [105].

Conclusions

Multifactorial cellular dysfunction is associated with iron dyshomeostasis in central nervous system resulting in iron accumulation. Mitochondrial dysfunction, protein misfolding and aggregation, autophagic-lysosomal dysfunction, neuroinflammation and ferroptosis are usually involved . Advances in MRI imaging techniques for specific detection of iron holds a great promise in future to consider iron as a biomarker for preclinical stages of neurodegeneration. Moreover, CSF levels of iron related proteins may aid in early diagnosis of sporadic neurodegenerative diseases in the future, like CSF ferritin in AD. However lot of questions has to be addressed in future regarding iron metabolism in human body right from brain sensing to different cells and pathways regulating iron content at intercellular level.

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Bibliography

- Lane DJR., et al. "Iron and Alzheimer's Disease: An Update on Emerging Mechanisms". Journal of Alzheimer's Disease 64.1 (2018): S379-S395.
- Ashraf A., et al. "The aging of iron man". Frontiers in Aging Neuroscience 10 (2018): 65.
- 3. Chang YZ. "Brain Iron Metabolism and CNS Diseases". *Advances in Experimental Medicine and Biology* (2019).
- Belaidi AA and Bush AI. "Iron neurochemistry in Alzheimer's disease and Parkinson's disease: targets for therapeutics". *Journal of Neurochemistry* 139 (2016): 179-197.

- 5. Duck KA., *et al.* "A role for sex and a common HFE gene variant in brain iron uptake". *Journal of Cerebral Blood Flow and Metabolism* 38 (2018): 540-548.
- Mccarthy RC and Kosman DJ. "Iron transport across the bloodbrain barrier: development, neurovascular regulation and cerebral amyloid angiopathy". *Cellular and Molecular Life Sciences* 72 (2015): 709-727.
- Rouault TA., *et al.* "Brain iron homeostasis, the choroid plexus, and localization of iron transport proteins". *Metabolic Brain Disease* 24 (2009): 673-684.
- 8. Chiou B., *et al.* "Endothelial cells are critical regulators of iron transport in a modelof the human blood-brain barrier". *Journal of Cerebral Blood Flow and Metabolism* (2018).
- Rudisill SS., *et al.* "Iron deficiency reduces synapse formation in the drosophila clock circuit". *Biological Trace Element Research* 189 (2019): 241-250.
- Khan AI., *et al.* "Iron transport kinetics through blood-brain barrier endothelial cells". *Biochimica et Biophysica Acta* 1862 (2018):1168-1179.
- 11. Leitner DF and Connor JR. "Functional roles of transferrin in the brain". *Biochimica et Biophysica Acta* 1820 (2012): 393-402.
- 12. Qian ZM and Shen X. "Brain iron transport and neurodegeneration". *Trends in Molecular Medicine* 7 (2001): 103-108.
- 13. Burkhart A., *et al.* "Expression of iron-related proteins at the neurovascular unit supports reduction and reoxidation of iron for transport through the blood-brain barrier". *Molecular Neurobiology* 53 (2016): 7237-7253.
- 14. Chiou B., *et al.* "Semaphorin 4A and H-ferritin utilize Tim-1 on human oligodendrocytes: a novel neuro-immune axis". *Glia* 66 (2018): 1317-1330.
- 15. Fishman J., *et al.* "Receptor mediated transcytosis of transferrin across the blood-brain barrier". *Journal of Neuroscience Research* 18 (1987): 299-304.
- 16. Leveugle B., *et al.* "Cellular distribution of the iron-binding protein lactotransferrin in the mesencephalon of Parkinson's disease cases". *Acta Neuropathologica* 91 (1996): 566-572.
- 17. Moroo I., *et al.* "Identification of a novel route of iron transcytosis across the mammalian blood-brain barrier". *Microcirculation* 10 (2003): 457-462.

- Gunshin H., *et al.* "Cloning and characterization of a mammalian proton-coupled metal-ion transporter". *Nature* 388 (1997): 482-488.
- 19. Vargas JD., *et al.* "Stromal cell-derived receptor 2 and cytochrome b561 are functional ferric reductases". *Biochimica et Biophysica Acta* 1651 (2003): 116-123.
- Wang X., *et al.* "Efflux of iron from the cerebrospinal fluid to the blood at the blood-CSF barrier: effect of manganese exposure". *Experimental Biology and Medicine (Maywood)* 233 (2008b): 1561-1571.
- Unger EL., *et al.* "Diurnal cycle influences peripheral and brain iron levels in mice". *Journal of Applied Physiology* 106 (2009): 187-193.
- 22. Boland B., *et al.* "Promoting theclearance of neurotoxic proteins in neurodegenerative disorders of ageing". *Nature Reviews Drug Discovery* 17 (2018): 660-688.
- 23. Trevaskis NL., *et al.* "From sewer to savior targeting the lymphatic system to promote drug exposure and activity". *Nature Reviews Drug Discovery* 14 (2015): 781-803.
- 24. Moos T. "Brain iron homeostasis". *Danish Medical Bulletin* 49 (2002): 279-301.
- 25. Moos T. "Increased accumulation of transferrin by motor neurons of the mouse mutant progressive motor neuronopathy (pmn/pmn)". *Journal of Neurocytology* 24 (1995): 389-398.
- 26. Knutson MD. "Non-transferrin-bound iron transporters". *Free Radical Biology and Medicine* 133 (2019): 101-111.
- Urrutia P., *et al.* "Inflammation alters the expression of DMT1, FPN1 and hepcidin, and it causes iron accumulation in central nervous system cells". *Journal of Neurochemistry* 126 (2013): 541-549.
- 28. Tripathi AK., *et al.* "Prion protein functions as a ferrireductase partner for ZIP14 and DMT1". *Free Radical Biology and Medicine* 84 (2015): 322-330.
- 29. Xu M., *et al.* "Differential regulation of estrogen in iron metabolism in astrocytes and neurons". *Journal of Cellular Physiology* 234 (2019): 4232-4242.
- Ji C., *et al.* "The ferroxidase hephaestin but not amyloid precursor protein is required for ferroportin-supported iron efflux in primary hippocampal neurons". *Cellular and Molecular Neurobiology* 38 (2018): 941-954.

- 31. Jeong SY and David S. "Age-related changes in iron homeostasis and cell death in the cerebellum of ceruloplasmin-deficient mice". *Journal of Neuroscience* 26 (2006): 9810-9819.
- 32. Zarruk JG., *et al.* "Expression of iron homeostasis proteins in the spinal cord in experimental autoimmune encephalomyelitis and their implications for iron accumulation". *Neurobiology of Disease* 81 (2015): 93-107.
- Helgudottir SS., *et al.* "Hepcidin mediates transcriptional changes in ferroportin mRNA in differentiated neuronal-like PC12 cells subjected to iron challenge". *Molecular Neurobiology* 56 (2019): 2362-2374.
- Yanatori I., *et al.* "Iron export through the transporter ferroportin 1 is modulated by the iron chaperone PCBP2". *Journal of Biological Chemistry* 291 (2016): 17303-17318.
- Loeffler DA., et al. "Ceruloplasmin immunoreactivity in neurodegenerative disorders". Free Radical Research 35 (2001): 111-118.
- Goodman L. "Alzheimer's disease; A clinico-pathologic analysis of twenty- three cases with a theory on pathogenesis". *The Journal of Nervous and Mental Disease* 118 (1953): 97-130.
- Smith MA., *et al.* "Iron accumulation in Alzheimer disease is a source of redox-generated free radicals". *Proceedings of the National Academy of Sciences of the United States of America* 94 (1997): 9866-9868.
- Collingwood JF., *et al.* "In situ characterization and mapping of iron compounds in Alzheimer's disease tissue". *Journal of Alzheimer's Disease* 7 (2005): 267-272.
- Bartzokis G and Tishler TA. "MRI evaluation of basal ganglia ferritin iron and neurotoxicity in Alzheimer's and Huntingon's disease". *Cellular and Molecular Biology* 46 (2000): 821-833.
- 40. Lei P., *et al.* "Tau deficiency induces parkinsonism with dementia by impairing APP-mediated iron export". *Nature Medicine* 18 (2012): 291-295.
- 41. Lei P., *et al.* "Tau protein: relevance to Parkinson's disease". *The International Journal of Biochemistry and Cell Biology* 42 (2010):1775-1778.

- Islam K and Levy E. "Carboxyl-terminal fragments of betaamyloid precursor protein bind to microtubules and the associated protein tau". *The American Journal of Pathology* 151 (1997): 265-271.
- 43. Hirsch EC., *et al.* "Iron and aluminum increase in the substantia nigra of patients with Parkinson's disease: an X-ray microanalysis". *Journal of Neurochemistry* 56 (1991): 446-451.
- Jellinger K., *et al.* "Brain iron and ferritin in Parkinson's and Alzheimer's diseases". *Journal of Neural Transmission - Parkinson s Disease and Dementia Section* 2 (1990): 327-340.
- 45. Li WJ., *et al.* "Dose- and time-dependent alpha synuclein aggregation induced by ferric iron in SK-N-SH cells". *Neuroscience Bulletin* 26 (2010): 205-210.
- Febbraro F., *et al.* "Alpha-synuclein expression is modulated at the translational level by iron". *Neuro Report* 23 (2012): 576-580.
- Rocha EM., et al. "Alpha-synuclein: pathology, mitochondrial dysfunction and neuroinflammation in Parkinson's disease". Neurobiology of Disease 109 (2017): 249-257.
- Wakabayashi K., *et al.* "The Lewy body in Parkinson's disease: molecules implicated in the formation and degradation of alpha-synuclein aggregates". *Neuropathology* 27 (2007): 494-506.
- 49. Salazar J., *et al.* "Divalent metal transporter 1 (DMT1) contributes to neurodegeneration in animal models of Parkinson's disease". *Proceedings of the National Academy of Sciences of the United States of America* 105 (2008): 18578-18583.
- Liu C., *et al.* "S-nitrosylation of divalent metal transporter 1 enhances iron uptake to mediate loss of dopaminergic neurons and motoric deficit". *The Journal of Neuroscience* 38 (2018): 8364-8377.
- 51. Bonn D. "Pumping iron in Parkinson's disease". *Lancet* 347 (1996): 1614.
- Fisher J., *et al.* "Ferritin: a novel mechanism for delivery of iron to the brain and other organs". *American Journal of Physiology* 293 (2007): C641-C649.
- Iancu TC. "Ultrastructural aspects of iron storage, transport and metabolism". *Journal of Neural Transmission (Vienna)* 118 (2011): 329-335.

- 54. Xu H., *et al.* "Activation of NMDA receptors mediated iron accumulation via modulating iron transporters in Parkinson's disease". *FASEB Journal* 13 (2018).
- 55. Lumsden AL., *et al.* "Huntingtin-deficient zebrafish exhibit defects in iron utilization and development". *Human Molecular Genetics* 16.16 (2007): 1905-1920.
- 56. Agrawal S., *et al.* "Brain mitochondrial iron accumulates in Huntington's disease, mediates mitochondrial dysfunction, and can be removed pharmacologically". *Free Radical Biology and Medicine* 120 (2018): 317-329.
- Domínguez JFD., *et al.* "Iron accumulation in the basal ganglia in Huntington's disease: cross-sectional data from the IMAGE-HD study". *Journal of Neurology, Neurosurgery, and Psychiatry* 87.5 (2016): 545-549.
- Van Bergen JMG., et al. "Quantitative susceptibility mapping suggests altered brain iron in premanifest huntington disease". American Journal of Neuroradiology 37.5 (2016): 789-796.
- 59. Fox JH., *et al.* "Mechanisms of copper ion mediated Huntington's disease progression". *PLoS One* 2.3 (2007): e334.
- Hogarth P. "Neurodegeneration with brain iron accumulation: diagnosis and management". *Journal of Movement Disorders* 8 (2015): 1-13.
- Zhou B., *et al.* "A novel pantothenate kinase gene (PANK2) is defective in Hallervorden-Spatz syndrome". *Nature Genetics* 28 (2001): 345-349.
- 62. Hayflick SJ and Hogarth P. "As iron goes, so goes disease?" *Haematologica* 96 (2011): 1571-1572.
- 63. Haack TB., *et al.* "Exome sequencing reveals de novo WDR45 mutations causing a phenotypically distinct, X-linked dominant form of NBIA". *American Journal of Human Genetics* 91 (2012): 1144-1149.
- Saitsu H., *et al.* "De novo mutations in the autophagy gene WDR45 cause static encephalopathy of childhood with neurodegeneration in adulthood". *Nature Genetics* 45 (2013): 445-449.
- 65. Hametner S., *et al.* "Iron and neurodegeneration in the multiple sclerosis brain". *Annals of Neurology* 74 (2013): 848-861.

- Haider L., *et al.* "Multiple sclerosis deep grey matter: the relation between demyelination, neurodegeneration, inflammation and iron". *Journal of Neurology, Neurosurgery, and Psychiatry* (2014).
- Walsh A., *et al.* "Multiple sclerosis: validation of MR imaging for quantification and detection of iron". *Radiology* 267 (2013): 531-542.
- Zheng W., et al. "Measuring iron in the brain using quantitative susceptibility mapping and X- ray fluorescence imaging". Neuroimage 78 (2013): 68-74.
- 69. Yao B., *et al.* "Chronic multiple sclerosis lesions: characterization with high- strength MR imaging". *Radiology* 262 (2012): 206-215.
- Bagnato F., *et al.* "Tracking iron in multiple sclerosis: a combined imaging and histopathological study at 7 Tesla". *Brain* 134 (2011): 3602-3615.
- Mahad D., *et al.* "Mitochondrial defects in acute multiple sclerosis lesions". *Brain* 131 (2008): 1722-1735.
- 72. Trapp BD and Stys PK. "Virtual hypoxia and chronic necrosis of demyelinated axons in multiple sclerosis". *The Lancet Neurology* 8 (2009): 280-291.
- 73. Eaton JW and Qian M. "Molecular bases of cellular iron toxicity". *Free Radical Biology and Medicine* 32 (2002): 833-840.
- Cui L., *et al.* "Oligodendrocyte progenitor cell susceptibility to injury in multiple sclerosis". *The American Journal of Pathology* 183 (2013): 516-525.
- 75. Hametner S., *et al.* "Iron and neurodegeneration in the multiple sclerosis brain". *Annals of Neurology* 74 (2013): 848-861.
- 76. Harris ZL., et al. "Aceruloplasminemia:molecular characterization of this disorder of iron metabolism". Proceedings of the National Academy of Sciences of the United States of America 92 (1995): 2539-2543.
- 77. Miyajima H., *et al.* "Aceruloplasminemia, an inherited disorder of iron metabolism". *Biometals* 16 (2003): 205-213.
- Di Patti MC., *et al.* "Dominant mutants of ceruloplasmin impair the copper loading machinery in aceruloplasminemia". *Journal* of Biological Chemistry 284 (2009): 4545-4554.

- 79. Kono S., *et al.* "Cys-881 is essential for the trafficking and secretion of truncated mutant ceruloplasmin in aceruloplasminemia". *Journal of Hepatology* 47 (2007): 844-850.
- Kono S., *et al.* "Biological effects of mutant ceruloplasmin on hepcidin-mediated internalization of ferroportin". *Biochimica et Biophysica Acta* 1802 (2010): 968-975.
- 81. Curtis AR., *et al.* "Mutation in the gene encoding ferritin light polypeptide causes dominant adult-onset basal ganglia disease". *Nature Genetics* 28 (2001): 350-354.
- Barbeito AG., *et al.* "Abnormal iron metabolism and oxidative stress in mice expressing a mutant form of the ferritin light polypeptide gene". *Journal of Neurochemistry* 109 (2009): 1067-1078.
- 83. Vidal R., *et al.* "Expression of a mutant form of the ferritin light chain gene induces neurodegeneration and iron overload in transgenic mice". *Journal of Neurochemistry* 28 (2008): 60-67.
- Cozzi A., *et al.* "Oxidative stress and cell death in cells expressing L-ferritin variants causing neuroferritinopathy". *Neurobiology of Disease* 37 (2010): 77-85.
- 85. Sanchez-Casis G., *et al.* "Pathology of the heart in Friedreich's ataxia:review of the literature and report of one case". *The Canadian Journal of Neurological Sciences. Le Journal Canadien des Sciences Neurologiques* 3 (1976): 349-354.
- Rotig A., *et al.* "Aconitase and mitochondrial iron-sulphur protein deficiency in Friedreich ataxia". *Nature Genetics* 17 (1997): 215-217.
- Puccio H., *et al.* "Mouse models for Friedreich ataxia exhibit cardiomyopathy, sensory nerve defect and Fe–S enzyme deficiency followed by intramitochondrial iron deposits". *Nature Genetics* 27 (2001): 181-186.
- Garcia-Yebenes I., *et al.* "Iron overload, measured as serum ferritin, increases brain damage induced by focal ischemia and early reperfusion". *Neurochemistry International* 61 (2012): 1364-1369.
- 89. Lipscomb DC., *et al.* "Low molecular weight iron in cerebral ischemic acidosis in vivo". *Stroke* 29 (1998): 487-492.
- 90. Selim MH and Ratan RR. "The role of iron neurotoxicity in ischemic stroke". *Ageing Research Reviews* 3 (2004): 345-353.

- 91. Hua Y., *et al.* "Brain injury after intracerebral hemorrhage: the role of thrombin and iron". *Stroke* 38 (2007): 759-762.
- 92. Wu H., *et al*. "Iron toxicity in mice with collagenase-induced intracerebral haemorrhage". *Journal of Cerebral Blood Flow and Metabolism* 31 (2011): 1243-1250.
- Garton T., et al. "Brain iron overload following intracranial haemorrhage". Stroke and Vascular Neurology 1 (2016): 172-184.
- Veyrat-Durebex C., *et al.* "Iron metabolism disturbance in a French cohort of ALS patients". *BioMed Research International* (2014): 485723.
- 95. Yu J., *et al.* "Serum ferritin is a candidate biomarker of disease aggravation in amyotrophic lateral sclerosis". *Biomedical Reports* 9 (2018): 333-338.
- Kokic AN., et al. "Biotransformation of nitric oxide in the cerebrospinal fluid of amyotrophic lateral sclerosis patients". Redox Report 10 (2005): 265-270.
- 97. Hozumi I., *et al.* "Patterns of levels of biological metals in CSF differ among neurodegenerative diseases". *Journal of the Neurological Sciences* 303 (2011): 95-99.
- 98. Ignjatovic A., *et al.* "Inappropriately chelated iron in the cerebrospinal fluid of amyotrophic lateral sclerosis patients". *Amyotrophic Lateral Sclerosis* 13 (2012): 357-362.
- Yasui M., *et al.* "Concentrations of zinc and iron in the brains of Guamanian patients with amyotrophic lateral sclerosis and parkinsonism-dementia". *Neurotoxicology* 14 (1993): 445-450.
- 100. Ince PG., *et al.* "Iron, selenium and glutathione peroxidase activity are elevated in sporadic motor neuron disease". *Neuroscience Letters* 182 (1994): 87-90.
- 101. Kasarskis EJ., *et al.* "Aluminum, calcium, and iron in the spinal cord of patients with sporadic amyotrophic lateral sclerosis using laser microprobe mass spectroscopy: a preliminary study". *Journal of the Neurological Sciences* 130 (1995): 203-208.
- 102. Markesbery WR., *et al.* "Neutron activation analysis of trace elements in motor neuron disease spinal cord". *Neurodegeneration* 4 (1995): 383-390.

60

- 103. Qian ZM and Wang Q. "Expression of iron transport proteins and excessive iron accumulation in the brain in neurodegenerative disorders". *Brain Research Reviews* 27 (1998):257-267.
- 104. Chen L., *et al.* "Ablation of the ferroptosis inhibitor glutathione peroxidase 4 in neurons results in rapid motor neuron degeneration and paralysis". *Journal of Biological Chemistry* 290 (2015): 28097-28106.
- 105. Puy V., *et al.* "Predominantrole of microglia in brain iron retention in Sanfilippo syndrome, a pediatric neurodegenerative disease". *Glia* 66 (2018): 1709-1723.

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