



In vitro Antiviral Activity of *Bacopa monnieri* (L.) Wettst Roots Methanolic Extract Against Dengue Virus Type 2

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Abstract

Dengue is the most widely spread arthropod-transmitted viral diseases of human beings, approximately 390 million people are getting symptomatic infections annually and more than 2 billion people are living in risk conditions. Specified antiviral drug has not approved yet against dengue and in some countries restricted introduction of dengue vaccine is the only source. The only there is only limited introduction of a dengue vaccine in some countries. *Bacopa monnieri* (L.) Wettst plant frequently called as Brahmi and water hyssop, belongs to the family of Plantaginaceae. *Bacopa monnieri* (L.) Wettst is used as a medicine to cure various conditions including infections. In this study, determination of maximum non-toxic dose (MNTD) of the *Bacopa monnieri* (L.) Wettst was done against Vero E6 cells under *in vitro* conditions with the methanolic extract. The antiviral assay of *Bacopa monnieri* (L.) Wettst based on cytopathic effects (CPE) represented by degree of inhibition against DENV serotype 2 strain. Results showed that *Bacopa monnieri* (L.) Wettst had significant anti-dengue activity in Vero E6 cell lines by lowering the levels of cellular infection as well as virus output. It is done with the 50% effective concentrations for DENV 2 for Vero E6 cell lines. These results of this study by using MTT assay method, in which the Percentage of inhibition in the presence of *Bacopa monnieri* methanolic extract for Anti DENV-2 activity was increased dose dependent manner from 1.56 to 100 µg in DENV-2 infected cell. The results indicated that *Bacopa monnieri* (L.) Wettst has a potentiality as an anti-viral agent against dengue infection for further development.

Keywords: Dengue Virus DENV 2; *Bacopa monnieri* (L.) Wettst; Plantaginaceae; Antiviral

Dengue is a mosquito-borne viral infection, found in tropical and sub-tropical climates worldwide, mostly in urban and semi-urban areas. The incidence of dengue has grown dramatically around the world in recent decades. A huge majority of cases are asymptomatic or mild and therefore the true dengue cases are under-reported. Numerous cases are also misidentified as other febrile illnesses [1]. Some studies revealed approximately 390 million dengue virus infections are observing annually, out of which 96 million

have distinct symptoms clinically [2]. Another study on the ubiquity of dengue infection is about 3.9 billion people are at a risk of infection with dengue viruses. In spite of a risk of infection surviving in 129 countries [3], the real implication occurred about 70% in Asia [2]. According to WHO the dengue cases have raised over 8 folds over the last 2 decades. Deaths happened due to dengue infections have raised from 960 to 4032 between the year 2000 and 2015 [3]. Presently, there is no particular treatment for dengue infection. However, significant medical treatment could save the patient's

lives [4]. Clinical manifestations of dengue infection ranges from asymptomatic: Dengue Fever (DF), Dengue Hemorrhagic Fever (DHF), eventually Dengue Shock Syndrome (DSS) [5]. The preventive measures for diminishing of the dengue infection are mostly vector control and suggested dengue vaccine [6]. Research says that particular dengue vaccine shows some promising effect to prevent severe dengue cases although further study is required [4]. For the development of a novel anti-viral agent invitro model experiments are using which is a main issue. The cell substrate requirements for virus replication restricting the screening of novel compounds. Due to this reason, only a few drugs have raised as efficient antiviral drugs [7]. Alternatively, to develop a novel anti-viral drug for dengue various medicinal plant extracts are going to be investigate [8]. Few phytochemical compounds such as flavonoid, phenolics, and terpenoid which would be extracted from natural products were suggested to have an antiviral activity against dengue [9,10]. *Bacopa monnieri* (L.) Wettst plant frequently called as Brahmi and water hyssop, belongs to the family Plantaginaceae. It is used traditionally in Asia to treat malaria, headache, diarrheal infections and for the memory enhancement [11-13]. Two most important flavonoids include luteolin and apigenin were identified in *Bacopa monnieri* plant [14]. In this study, we move forward to evaluate the antiviral activity of *Bacopa monnieri* against DENV- 2 virus. The control over fever in dengue infection was done by using various synthetic types of drugs including aspirin, paracetamol. So many adverse effects are there by taking these synthetic types of drugs. Therefore, herbal medicines are used in the form of alternative therapy as they possess lesser side effects and more availability.

Materials and methods

Collection of plant material and its authentication

Roots of *Bacopa monnieri* (L.) Wettst plant was collected from the area of Andhra University, Visakhapatnam. The authentication was done by Dr. Padal, Taxonomist, Department of Botany, Andhra University, Andhra Pradesh. The roots were sorted out, cleaned. These roots were chopped and dried at room temperature. This can be grounded into a coarse powder.

Methanol extract preparation

The coarse powder of root material was extracted with methanol by using soxhlation process and gets filtered. By using a rotary evaporator, the extract was concentrated. The concentrated mass was placed in air tight container and kept in a desiccator for further studies.

Preparation of extract for cytotoxicity and antiviral assay

For the cytotoxicity and antiviral assays, a stock solution was prepared by dissolving 1.0 g of methanolic extract of *Bacopa monnieri* in 100 mL of dimethyl sulfoxide (DMSO) (Sigma Aldrich, USA). The stock solution was filter sterilised (0.20 µm pore, Minisart, Catalog No. 754004722) and further diluted with culture medium to the desired concentration for the assays. In this study Seven Concentrations ranging from 100µg/0.1ml, 50 µg/0.1ml, 25µg/0.1ml, 12.50 µg/0.1ml, 6.25 µg/0.1ml, 3.12 µg/0.1ml, and 1.56 µg/0.1 ml were chosen.

Preparation of medium

Powdered Dulbecco's Modified Eagle's Medium (DMEM) (GIBCO, UK) was used in this study. A total of 3.7 g of sodium bicarbonate was added, dissolved with 1 L of ultrapure water and the pH of the medium was adjusted to 7.0. The medium was then filter sterilised using 0.22 µm PES membrane filter (TPP, Switzerland) under vacuum condition.

Cell line

Vero E6 cell lines was procured from National Centre for Cell Sciences (NCCS), Pune, India and maintained in Dulbecco's Modified Eagle's Medium (DMEM) with 10% v/v Foetal bovine serum (FBS), L-Glutamine, Sodium bicarbonate, HEPES (2-[4-(2-Hydroxyethyl)1-piperziny] ethane sulphonic acid) buffer, Penicillin (TC020-100U/ml), Streptomycin (TC035- 100µg/ml) and Amphotericin-B (TC019-2.5µg/ml).

Maintenance and sub culturing of cells

The cryopreserved Vero E6 cells were rapidly thawed at 37°C in a water bath. Cells were transferred carefully to 25 cm² tissue culture flasks (Corning, USA) containing 4 mL of DMEM with 10% of Fetal Bovine Serum (FBS) (GIBCO, South America). Cells were then incubated at 37°C with 5% CO₂ for 2 to 3 days until confluent. At 80-100% confluency, the cells were subcultured. The subculturing process was initiated by removing the used medium followed by rinsing the cells twice with Phosphate buffer solution (PBS) (MP Biomedical, France). Then, 1 mL of 0.25% trypsin-EDTA (GIBCO, Canada) was added and incubated for 5 to 10 minutes at 37°C. After trypsinisation, 1 mL of fresh medium with 10% FBS was added and mixed. The mixture was then centrifuged for 10 minutes at 1500 rpm. The supernatant was discarded while the pellet was resuspended with 2 mL of fresh medium and re-distributed into new tissue culture flask for further maintenance.

Counting of cells

The cells were counted using a hemocytometer. The cell suspension was mixed gently and an aliquot to the trypan blue solution was added (100ul cell suspension: 100 ul dye). The sample was mixed thoroughly and was pipetted into the junction between the counting chamber and the cover slip of hemocytometer. It was observed under light microscope. The viable and nonviable cells in both halves of the chamber were calculated.

In vitro cytotoxicity assay by MTT method

MTT (3-(4, 5-dimethylthiazol-2-yl)-2,5-diphenyl tetrazolium bromide (Himedia catalog no.TC191) was dissolved in PBS at 5 mg/ml and filtered to remove insoluble particles of MTT. After the incubation period, 20µl of MTT solution was added to all wells and plates were incubated at 37°C at 5% CO2 atmosphere for 4 hours. After 4 hours, 100 µl of DMSO was added to wells and incubated for 10 minutes. The plates were read on a Microplate reader, using a test wavelength of 540 nm [15].

Antiviral assay

DENV serotype 2 strain New Guinea C (NGC)

DENV-2 (M29095) strain was procured from National Institute of Virology (NIV) maintained in Virus Transport Medium (VTM) at 4°C. 0.1 ml of the VTM supernatant and maintenance medium was transferred on to the monolayer of DENV-2 cell line and incubated at 37°C for 96 hours. Complete cytopathic effect (CPE) in DENV-2 cell line was observed on the 4th day.

In vitro antiviral assay by MTT method

20µl of MTT solution [3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyl-2H-tetrazolium bromide] was added to all the wells. Plates were incubated at 37°C at 5% CO2 atmosphere for 4 hours. After 4 hours 100 µl of DMSO was added to wells and incubated for 10 minutes. At a wavelength of 540 nm the plates were read by using a Microplate reader. A gross absence of cytopathic effect was inferred by intact of cell layer, without distortion in the morphology of cells or nucleus and syncytial cells in comparison with the control rows. Hence wells with or without CPE were marked. Through Inverted Tissue Culture Microscope, an important observation of the incubated wells, both experimental and control were recorded by microphotography. The fifty percentage of inhibitory effect will be considered for antiviral study.

Statistical analysis

The *in vitro* cytotoxicity and antiviral activity of selected plant extracts was assessed by statistical analysis data comprising standard mean and standard deviation. P values were calculated using Graph Pad Prism 7 and graph was plotted based on the values.

Results and Discussion

In vitro cytotoxicity assay

The evaluation of cytotoxic potential of methanol extract of selected plants was investigated using the MTT assay.

In vitro cytotoxicity assay for methanol extract of Bacopa monnieri

The methanol extract of *Bacopa monnieri* was found to be toxic free from concentrations of 1.56 µg/0.1ml to 25 µg/0.1ml and concentrations of 50 and 100 µg/0.1ml were found to toxic in the Vero E6 cell line (Table 1). The triplicate optical density values of the MTT assay was represented in table 2. Statistical analysis and graphical representation of cytotoxicity of methanol extract of *Bacopa monnieri* showed that at 25 µg/0.1ml concentration found to be highly significant (Table 3 and Figure 1). The percentage of cell viability varied from 99% to 78% (Figure 2), thus it is proposed that there is no remarkable cytotoxic activity effect on the Vero E6 cell lines upto 50 µg/0.1ml represented (Table 4).

S. No	Concentrations (µg/0.1ml)	Cytotoxicity	
		<i>Bacopa monnieri</i>	Cell control
1.	1.56	-	-
2.	3.12	-	-
3.	6.25	-	-
4.	12.5	-	-
5.	25	-	-
6.	50	+	-
7.	100	+	-

Table 1: Cytotoxicity profile of methanolic extract of selected plant

(+) → Presence of cytotoxicity

(-) → Absence of cytotoxicity.

Concentrations (µg/0.1ml)	Absorbance at 540nm					
	<i>Bacopa monnieri</i>			Cell control		
1.56	1.215	1.213	1.214	1.226	1.22	1.226
3.12	1.213	1.21	1.212	1.223	1.216	1.223
6.25	1.208	1.203	1.206	1.218	1.211	1.218
12.5	1.192	1.188	1.19	1.223	1.207	1.223
25	1.183	1.18	1.181	1.214	1.204	1.214
50	1.007	1.002	1.005	1.22	1.215	1.22
100	0.949	0.943	0.945	1.216	1.206	1.211

Table 2: Cytotoxicity studies of methanolic extract of *Bacopa monnieri* by MTT assay.

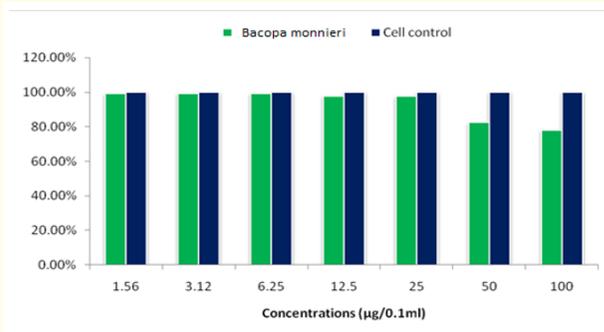


Figure 2: Percentage of viable cells in the presence of selected methanolic extract *Bacopa monnieri* for cytotoxicity assay.

Concentrations (µg/0.1ml)	<i>Bacopa monnieri</i>	Cell control
1.56	1.214 ± 0.0006	1.224 ± 0.002
3.12	1.212 ± 0.0009	1.221 ± 0.0023
6.25	1.206 ± 0.0015	1.216 ± 0.0023
12.5	1.19 ± 0.0012	1.218 ± 0.0053
25	1.181 ± 0.0009	1.211 ± 0.0033
50	1.005 ± 0.0015	1.218 ± 0.0017
100	0.946 ± 0.0018	1.211 ± 0.0029
P Value	***	***

Table 3: Statistical analysis of cytotoxicity of selected methanolic extract.

Concentrations (µg/0.1ml)	<i>Bacopa monnieri</i>	Cell control
1.56	99.18%	100%
3.12	99.26%	100%
6.25	99.18%	100%
12.5	97.73%	100%
25	97.58%	100%
50	82.46%	100%
100	78.09%	100%

Table 4: Percentage of viable cells in the presence of selected methanolic extract for cytotoxicity assay.

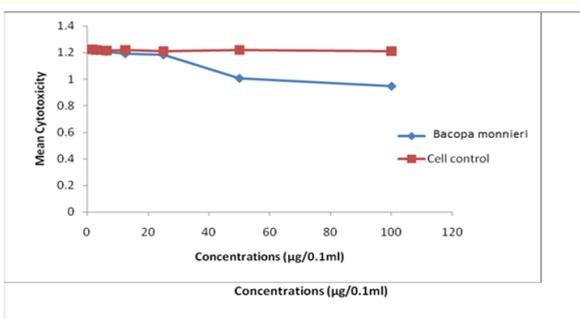


Figure 1: Cytotoxicity of selected methanolic extract of *Bacopa monnieri*.

In vitro antiviral assay

The evaluation of antiviral potential of methanol extract of selected plants was investigated using the MTT assay.

In vitro antiviral assay for methanol extract of *Bacopa monnieri*

In vitro anti DENV-2 activity was performed with extract of methanol from *Bacopa monnieri*. The methanol extract showed inhibitory activity from 50 µg/0.1ml and 12.5 µg/0.1ml concentrations (Table 5). The triplicate optical density values of MTT assay of anti DENV-2 activity was obtained (Table 6). Statistical analysis and graphical representation of antiviral activity of methanolic extract of *Bacopa monnieri* was represented table 7 and figure 3. The percentage of inhibition was represented table 8 and figure 4.

Concentrations (µg/0.1ml)	Cytopathic Effect	Cell control	Virus control
	<i>Bacopa monnieri</i>		
1.56	+	-	+
3.12	+	-	+
6.25	+	-	+
12.5	-	-	+
25	-	-	+
50	-	-	+
100	**	-	+

Table 5: Antiviral activity of methanolic extract of selected plant.

(+) → Presence of cytopathic effect

(-) → Absence of cytopathic effect

(**) → Not Performed due to cytotoxicity.

Concentrations (µg/0.1ml)	Absorbance at 540nm					
	<i>Bacopa monnieri</i>			Cell control		
1.56	0.198	0.184	0.271	1.248	1.246	1.243
3.12	0.369	0.377	0.34	1.235	1.239	1.237
6.25	0.547	0.568	0.558	1.247	1.242	1.245
12.5	0.699	0.681	0.689	1.239	1.236	1.238
25	0.728	0.725	0.781	1.234	1.238	1.248
50	NP	NP	NP	1.246	1.246	1.235
100	NP	NP	NP	1.248	1.244	1.248

Table 6: Anti DENV-2 activity of methanolic extract of *Bacopa monnieri* by MTT assay.

Concentrations (µg/0.1ml)	<i>Bacopa monnieri</i>	Cell control
1.56	0.218 ± 0.0270	1.246 ± 0.0015
3.12	0.362 ± 0.0113	1.237 ± 0.0012
6.25	0.558 ± 0.0061	1.245 ± 0.0015
12.5	0.690 ± 0.0052	1.238 ± 0.0009
25	0.745 ± 0.0182	1.240 ± 0.0042
50	Not Performed	1.242 ± 0.0037
100	Not Performed	1.247 ± 0.0013
P Value	***	***

Table 7: Statistical analysis of antiviral activity of selected methanolic extract of *Bacopa monnieri*.

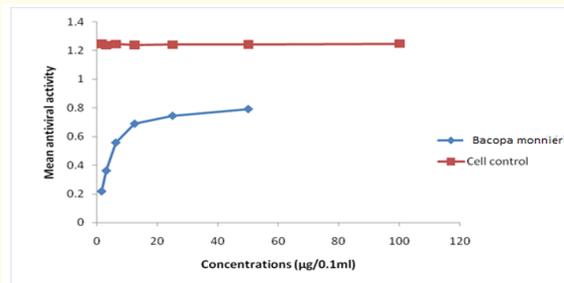


Figure 3: Antiviral activity of selected methanolic extract of *Bacopa monnieri*.

Concentrations (µg/0.1ml)	<i>Bacopa monnieri</i>	Cell control
1.56	17.47%	100%
3.12	29.26%	100%
6.25	44.80%	100%
12.5	55.72%	100%
25	60.05%	100%
50	67.35%	100%
100	Not Performed	100%

Table 8: Percentage of inhibition in the presence of selected methanolic extract for Anti DENV-2 activity.

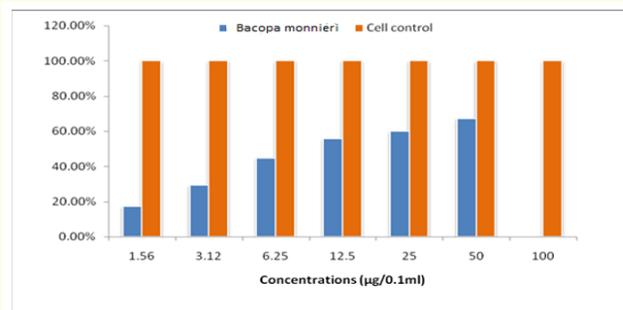


Figure 4: Percentage of inhibition in the presence of selected methanolic extract *Bacopa monnieri* for Anti DENV-2 activity.

Conclusion

The cytotoxicity as well as antiviral activity of *B. monnieri* roots extract exhibited significant ($p < 0.05$) reduction in DENV-2 infe-

ected cells and these results are compared to control. Without any cytotoxic effect the methanolic extract of *Bacopa monnieri* has antiviral effect against DENV-2. There is a need to study further to explore the IC50 values, CC50 values and mechanism of action regarding anti-viral activity of those plant extracts. Purification process as well as characterization of active compounds should be done furthermore to define antiviral DENV activity. The preliminary phytochemical screening of the methanolic extract of *Bacopa monnieri* plant has also revealed the presence of various phytochemicals such as flavonoids, alkaloids, proteins, steroids and carbohydrates. Hence, may be due to the presence of these phytochemicals the activity was possessed by this plant.

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