



Genotype-Phenotype Correlation in Spinal Muscular Atrophy

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Abstract

Spinal muscular atrophy (SMA) is an autosomal recessive neurodegenerative lower motor neuron disease with an incidence of 1/6000-1/10000 world-wide and characterised by symmetrical muscle weakness. Based on the age at onset and clinical severity, SMA classified clinically into the following types: SMA 0 (prenatal onset), SMA I (Werdnig-Hoffmann disease with onset before age 6 months), SMA II (Dubowitz disease with onset between age 6 and 18 months), SMA III (Kugelberg-Welander disease with onset after age 18 months), and SMA IV (onset after age 18 years). All types of SMA are the result of mutations in survival motor neuron (*SMN1*) gene and infantile type is the second leading genetic cause of death in infants. Although, mutations in *SMN1* gene are essential for pathogenesis of the SMA, copy number variation in *SMN2* gene and modification by some are genes such as *NAIP*, *SERF1A* and *PLS3* determine the age of onset and the severity of disease in its phenotypic continuum.

Keywords: SMA; SMN; NAIP; Spinal Muscular Atrophy; Werdnig-Hoffmann; Kugelberg-Welander

Introduction

Spinal muscular atrophy (SMA), with an incidence of 1/6000-1/10000 world-wide [1] is the most common childhood genetic lower motor neuron disease characterized by symmetrical muscle weakness with significant phenotype variability. Based on the age at onset and clinical severity, SMA classified clinically into the following types: SMA 0 (prenatal onset), SMA I (Werdnig-Hoffmann disease with onset before age 6 months), SMA II (Dubowitz disease with onset between age 6 and 18 months), SMA III (Kugelberg-Welander disease with onset after age 18 months), and SMA IV (onset after age 18 years). All types of SMAs are the result of mutation in the survival motor neuron 1 (*SMN1*) gene and the severity of SMAs related to the copy number of *SMN2* gene and modification with some other genes. SMN gene located on 5q11.2-q13.3 chromosomal region [2,3], and this region is a duplicated area of approximately 500 kb, where, at least four genes (SMN, NAIP, SERF1 and GTF2H2) are duplicated. Each of these duplicated genes has a telomeric and

a centromeric copy.

SMN gene has two homologue copies, *SMN1* (telomeric copy) and *SMN2* (centromeric copy), which differ by only eight nucleotides (five are intronic and three are exonic, located within exons 6, 7, and 8) [4-7].

One of the coding sequence of *SMN2* that differs from that of *SMN1* by a single nucleotide (840 C > T) results in alternative splicing of exon 7 [5], which consequently leads to reduced amount of full length transcripts [3], which is insufficient to counteract the development of SMA. So, SMA is caused by low levels of SMN protein rather than the complete absence of SMN [8,9]. Although, mutations in *SMN1* gene are essential for pathogenesis of the SMA, copy number variation in *SMN2* gene and modification by some are genes such as *NAIP*, *SERF1A* and *PLS3* determine the age of onset and the severity of disease in its phenotypic continuum.

SMN1 gene:

SMN1 gene was introduced as a candidate gene for SMA in 1995 [4,10,11]. Reduced levels of SMN result in degeneration of α motor neurons in the spinal cord, leading to muscle atrophy and weakness in SMA patients [12]. SMN is a housekeeping protein involved in small nuclear ribonucleoprotein biogenesis, neuromuscular junction formation, axonal growth, and transport of RNA along axons [13-15].

Approximately, 95% - 98% of all types of SMA patients show homozygous deletion of *SMN1* exon 7 [16,17]. Only 2 - 5% of SMA patients are compound heterozygous with a deletion of exon 7 and a point mutation [18-22]. Homozygous subtle mutations are very rare in patients with SMA [23,24].

SMN2 gene:

Depending on the copy number of the gene, *SMN2* produces a reduced amount of full length transcripts [3], which is insufficient to prevent the development of SMA. So, the severity of SMA is related to the reduced levels of SMN protein rather than the complete absence of SMN [8,9]. Approximately 5% of normal individuals do not carry the *SMN2* gene [4].

The number of *SMN2* copies (arranged in tandem in cis configuration on each chromosome) ranges from zero to five that can be detected using quantitative PCR and MLPA methods [25,26]. Copy number variation or gene dosage can also be determined via MAPH technique [27]. The presence of three or more copies of *SMN2* is associated with a milder phenotype [28-31]. Even, unaffected patients with a homozygous deletion of the *SMN1* gene, with four or five *SMN2* copies, have been reported [32].

As, there is a significant relationship between the clinical phenotype and *SMN2* copy number, *SMN2* can be considered as an important SMA-modifying gene [33]. A nucleotide variation in exon 7 of the *SMN2* gene (c.859G > C) has been described as a positive phenotype modifier in SMA patients, that is, found in SMA patients with a lower *SMN2* copy number than expected according to their phenotypes [34-36]. So, considering this nucleotide variation in the *SMN2* gene, it seems that all copies of the *SMN2* genes don't have a similar protective effect.

Other genes in the 5q13.2 region:

NAIP (neuronal apoptosis inhibitory protein) and *SERF1A* (small EDRK-rich factor 1A) genes have been suggested as possible SMA modifier genes. These genes are deleted in approximately 50% of the patients with severe SMA [10,33,37,38]. In a recent study, *NAIP* and *SERF1A* copy number showed a positive correlation with the SMA phenotype, where, *NAIP* genes was absent in nearly 73% of type I SMA and in only a few cases with type II and III disease, and *SERF1A* was absent in the 35% of type I SMA and in only one case with type II [39].

Higher frequency of homozygous deletion of *SMN1* gene in severe type I SMA suggests involvement of *NAIP* deletion in the SMA phenotype. Deletion in the *NAIP* gene can worsen the prognosis independent of the number of *SMN2* copy numbers [40-42]. *NAIP* gene is an apoptosis inhibitor, thus its deletion may result in the loss of spinal motor neurons.

Plastin 3 (PLS3) gene:

Recent documents revealed that *PLS3*, located on chromosome Xq23 and highly expressed in the spinal cord, can play a role as a positive modifier of SMA phenotype [43,44]. *PLS3* is a Ca^{2+} dependent F-actin-binding protein that plays an important role in axon development, cell polarity and migration [45,46].

PLS3 can fully protect against SMA in *SMN1*-deleted individuals carrying 3 - 4 *SMN2* copies, where, *SMN2* products is insufficient to counteract the development of SMA.

It has been revealed that in some rare families with unaffected homozygous *SMN1*-deleted females, the expression of *PLS3* was higher than in their affected counterparts. The protective modifier effect of *PLS3* may be due to its axon genesis role [47] that can rescue the axonal growth defects [48].

SMA type 0:

SMA Type 0 or congenital SMA (sometimes classified as SMA type Ia) is the most severe type with prenatal onset [49-51]. The presence of only one copy of *SMN2* has been described mostly in patients with congenital SMA or severe neonatal forms [52,53]. No patient has been reported with a homozygous deletion of both *SMN1* and *SMN2* genes, and this may be due to in utero lethality of this condition.

SMA type I (Werdnig-Hoffmann disease):

Type I SMA is a severe type which shows generalized muscle weakness and hypotonia with onset in the first six months of life. Patients with affected Type I SMA never sit without aid and generally die of respiratory failure before two years of age [54]. Type 0 or congenital SMA sometimes classified as SMA type Ia, where, the classical form of the disease with onset after the neonatal period considered as type Ib, and patients with head control as type Ic [49,55]. Approximately, all SMA type I patients have two copies of *SMN2* gene regardless of subtypes Ib and Ic [33,39].

SMA type II (Dubowitz disease):

The onset of symptoms in SMA type II occurs between 6 and 18 months. Patients can sit but unable to walk without aid [54]. Depending on the respiratory involvement and management of the complications they can reach adolescence and even adult age.

Most SMA type II patients have three copies of *SMN2* gene [33]; however, rarely SMA type II patient with only one copy of the *SMN2* gene has been reported [41].

SMA type III (Kugelberg-Welander):

Patients with Kugelberg-Welander disease or juvenile SMA are able to walk and the lifespan is generally not reduced [54]. SMA type III classified into two types of IIIa with onset before three years of age and IIIb with onset between three and 20 years of age. The probability of being able to walk after 10, 20 and 40 years of age is 73%, 44% and 34%, respectively, in SMA IIIa, and 97%, 89% and 67% in SMA IIIb [56,57].

SMA type III patients generally have three or four *SMN2* copies [33,39]. The presence of four copies of the *SMN2* gene is more frequent in type IIIb than in type IIIa SMA [58,39]. SMA type III patients with more than three *SMN2* copies show better motor function over time regardless of age at onset [30]. However, the influence of *SMN2* copy number is not strict, e.g., three *SMN2* copies have been detected in both SMA I and SMA III. One explanation may be that all *SMN2* copies are not functionally equivalent [59].

SMA type IV (Adult SMA):

SMA type IV is a less common form of the disease and its symptoms manifest between 20 and 30 years of age with a normal life expectancy [50,60,61]. The high copy number of *SMN2* in types IV SMA, generally more than three copies, can partially compensate for the absence of *SMN1* product [28], and more than four copies of *SMN2*, even 6 copies, are also reported in milder type or type IV SMA [25,58,62]. Since, general population has an average of one or two copies of *SMN2* gene, greater copy number in the mild form of SMA probably result from the conversion of *SMN1* into *SMN2* gene [63].

Phenotypic Discordances:

Usually, siblings affected with SMA are very similar in their clinical presentations, in terms of age at onset and the progression of disease. However, in rare cases, phenotypic discordances can be seen in the SMA patients, that is, individuals are asymptomatic or mildly affected despite carrying the same *SMN1* mutations as their affected siblings, which suggests the effect of genetic modifiers [47].

Phenotypic discordances have been reported between haploidentical siblings with milder forms in adulthood i.e. SMA type III. Patients with severe forms (type I and type II) tended to show fairly similar phenotypic presentation [32]. The Phenotypic discordances could be due to the presence of genetic phenotypic modifiers other than *SMN2* that may act in early life. For instance, *PLS3* has higher expression in unaffected *SMN1*-deleted individuals in comparison with their affected siblings [47]. *PLS3* is an important factor for the process of axonogenesis through increasing the level of F-actin, so defects in the axonogenesis may be the major cause in the pathogenesis of SMA [47].

Gender Effect:

The influence of gender on the phenotype of SMA remains unclear; however, it seems to play a role in the severity of disease. In a study on 1039 SMA patients, the overall ratio of females to males was F/M = 0.82, and the gender disproportion was higher for milder forms, that is, F/M ratio for SMA3b was 0.45 [41].

Milder forms of SMA, with the onset at the age of over 3 years, were seen approximately twice as frequently in males than females, and it is suggested that estrogens may play as a protective role in milder forms in females [41,64]. Also, asymptomatic cases with biallelic mutation of the *SMN1* gene have been reported more frequently in women than in men [41,65,66]. On the other hand, the more severe genotype, that is, *NAIP* gene deletion and the presence of two *SMN2* copies, was observed more frequently in female than males [41,67].

Summary

All types of SMA result from mutations in *SMN1* gene and its significant variations in the age of onset and the severity of clinical symptoms are due to modifier genes. Full-length product of *SMN1* is necessary for lower motor neuron function and loss of *SMN1* is essential to the pathogenesis of SMA, while *SMN2* copy number modifies the severity of phenotype. No correlation exists between the loss of *SMN1* exon 7 and the severity of disease, that is, the homozygous exon 7 deletion is observed with the same frequency in all phenotypes. It seemed that a large deletion including neighbouring genes such as *NAIP* and *SERF1A* cause the severe phenotypes of SMA. And also, *PLS3* gene, located on chromosome Xq23, can play a role as a positive modifier of SMA phenotype.

Bibliography

1. Sugarman EA, et al. "Pan-ethnic carrier screening and prenatal diagnosis for spinal muscular atrophy: clinical laboratory analysis of > 72,400 specimens". *European Journal of Human Genetics* 20.1 (2012): 27-32.
2. Melki J, et al. "Gene for chronic proximal spinal muscular atrophies maps to chromosome 5q". *Nature* 344.6268 (1990): 767-768.
3. Brzustowicz LM, et al. "Genetic mapping of chronic childhood-onset spinal muscular atrophy to chromosome 5q11.2-13.3". *Nature* 344.6266 (1990): 540-541.
4. Lefebvre S, et al. "Identification and characterization of a spinal muscular atrophy-determining gene". *Cell* 80.1 (1995): 155-165.
5. Burglen L, et al. "Structure and organization of the human survival motor neurone (SMN) gene". *Genomics* 32.3 (1996): 479-482.
6. Chen Q, et al. "Sequence of a 131-kb region of 5q13.1 containing the spinal muscular atrophy candidate genes SMN and NAIP". *Genomics* 48.1 (1998): 121-127.

7. Biros I and Forrest S. "Spinal muscular atrophy: untangling the knot?". *Journal of Medical Genetics* 36.1 (1999): 1-8.
8. Burghes AH and Beattie CE. "Spinal muscular atrophy: why do low levels of survival motor neuron protein make motor neurons sick?". *Nature Reviews Neuroscience* 10.8 (2009): 597-609.
9. Prior TW. "Perspectives and diagnostic considerations in spinal muscular atrophy". *Genetics in Medicine* 12.3 (2011): 145-152.
10. Roy N., et al. "The gene for neuronal apoptosis inhibitory protein is partially deleted in individuals with spinal muscular atrophy". *Cell* 80.1 (1995): 167-178.
11. Lewin B. "Genes for SMA: Multum in Parvo". *Cell* 80.1 (1995): 1-5.
12. Merlini L., et al. "Motor function-muscle strength relationship in Spinal muscular atrophy". *Muscle and Nerve* 29.4 (2004): 548-552.
13. McWhorter ML., et al. "Knockdown of the survival motor neuron (Smn) protein in zebrafish causes defects in motor axon outgrowth and pathfinding". *The Journal of Cell Biology* 162.5 (2003): 919-931.
14. Zhang HL., et al. "Active transport of the survival motor neuron protein and the role of exon-7 in cytoplasmic localization". *Journal of Neuroscience* 23.16 (2003): 6627-6637.
15. Carrel TL., et al. "Survival Motor Neuron Function in Motor Axons Is Independent of Functions Required for Small Nuclear Ribonucleoprotein Biogenesis". *The Journal of Neuroscience* 26.43 (2006): 11014-11022.
16. Scheffer H., et al. "SMA carrier testing-validation of hemizygous SMN exon 7 deletion test for the identification of proximal spinal muscular atrophy carriers and patients with a single allele deletion". *European Journal of Human Genetics* 8.2 (2000): 79-86.
17. Ogino S and Wilson RB. "Genetic testing and risk assessment for spinal muscular atrophy (SMA)". *Human Genetics* 111.6 (2002): 477-500.
18. Parsons DW., et al. "Intragenic telSMN mutations: frequency, distribution, evidence of a founder effect, and modification of the spinal muscular atrophy phenotype by cenSMN copy number". *American Journal of Human Genetics* 63.6 (1998): 1712-1723.
19. Wirth B. "An update of the mutation spectrum of the survival motor neuron gene (SMN1) in autosomal recessive spinal muscular atrophy". *Human Mutation* 15.3 (2000): 228-237.
20. Fraidakis MJ., et al. "Genotype-phenotype relationship in 2 SMA III patients with novel mutations in the Tudor domain". *Neurology* 78.8 (2012): 551-556.
21. Ganji H., et al. "Detection of Intragenic SMN1 Mutations in Spinal Muscular Atrophy Patients with a Single Copy of SMN1". *Journal of Child Neurology* 30.5 (2015): 558-562.
22. Zabnenkova VV., et al. *Russian Journal of Genetics* 51 (2015): 925.
23. Kirwin SM., et al. "A homozygous double mutation in SMN1: a complicated genetic diagnosis of SMA". *Molecular Genetics and Genomic Medicine* 1.2 (2013): 113-117.
24. Rad IA., et al. "Homozygous Point Mutation in a Patient with Spinal Muscular Atrophy Type 1". *Journal of Genetic Disorders and Genetic Reports* 5.3 (2016).
25. Arkblad EL., et al. "Multiplex ligation-dependent probe amplification improves diagnostics in spinal muscular atrophy". *Neuromuscular Disorders* 16.12 (2006): 830-838.
26. Scarciolla O., et al. "Spinal muscular atrophy genotyping by gene dosage using multiple ligation-dependent probe amplification". *Neurogenetics* 7.4 (2006): 269-276.
27. Armour JA., et al. "Gene dosage analysis by multiplex amplifiable probe hybridization". *Methods in Molecular Medicine* 92 (2004): 125-139.
28. Feldkotter M., et al. "Quantitative analyses of SMN1 and SMN2 based on real-time light Cyclor PCR: fast and highly reliable carrier testing and prediction of severity of spinal muscular atrophy". *The American Journal of Human Genetics* 70.2 (2002): 358-368.
29. Soler-Botija C., et al. "Implication of fetal SMN2 expression in type I SMA pathogenesis: protection or pathological gain of function?". *Journal of Neuropathology and Experimental Neurology* 64.3 (2005): 215-223.
30. Swoboda KJ., et al. "Natural history of denervation in SMA: relation to age, SMN2 copy number, and function". *Annals of Neurology* 57.5 (2005): 704-712.
31. Zheleznyakova GY., et al. "Genetic and expression studies of SMN2 gene in Russian patients with spinal muscular atrophy type II and III". *BMC Medical Genetics* 12 (2011): 96.
32. Jedrzejowska M., et al. "Unaffected patients with a homozygous absence of the SMN1 gene". *European Journal of Human Genetics* 16.8 (2008): 930-934.
33. Noguchi Y., et al. "Telomeric region of the spinal muscular atrophy locus is susceptible to structural variations". *Pediatric Neurology* 58 (2016): 83-89.
34. Prior TW., et al. "A positive modifier of spinal muscular atrophy in the SMN2 gene". *American Journal of Human Genetics* 85.3 (2009): 408-413.
35. Bernal S., et al. "The c.859G>C variant in the SMN2 gene is associated with types II and III SMA and originates from a common ancestor". *Journal of Medical Genetics* 47.9 (2010): 640-642.
36. Vezain M., et al. "A rare SMN2 variant in a previously unrecognized composite splicing regulatory element induces exon 7 inclusion and reduces the clinical severity of spinal muscular atrophy". *Human Mutation* 31.1 (2010): 1110-1125.

37. He J., *et al.* "Molecular analysis of SMN1, SMN2, NAIP, GTF2H2, and H4F5 genes in 157 Chinese patients with spinal muscular atrophy". *Gene* 518.2 (2013): 325-329.
38. Brkusanin M., *et al.* "Joint effect of the SMN2 and SERF1A genes on childhood-onset types of spinal muscular atrophy in Serbian patients". *Journal of Human Genetics* 60.11 (2015): 723-728.
39. Medrano S., *et al.* "Genotype-phenotype correlation of SMN locus genes in spinal muscular atrophy children from Argentina". *European Journal of paediatric Neurology* 20.6 (2016): 910-917.
40. Dastur RS., *et al.* "Correlation between deletion patterns of SMN and NAIP genes and the clinical features of spinal muscular atrophy in Indian patients". *Neurology India* 54.3 (2006): 255-259.
41. Jedrzejowska M., *et al.* "Phenotype modifiers of spinal muscular atrophy: the number of SMN2 gene copies, deletion in the NAIP gene and probably gender influence the course of the disease". *Acta Biochimica Polonica* 56.1 (2009): 103-108.
42. Ahn EJ., *et al.* "Genotype-Phenotype Correlation of SMN1 and NAIP Deletions in Korean Patients with Spinal Muscular Atrophy". *Journal of Clinical Neurology* (2016): 1-5.
43. Heesen L., *et al.* "Plastin 3 is upregulated in iPSC-derived motoneurons from asymptomatic SMN1-deleted individuals". *Cellular and Molecular Life Sciences* 73.10 (2015): 2089-2104.
44. Hosseinibarkooie SM., *et al.* "The Power of Human Protective Modifiers: PLS3 and CORO1C Unravel Impaired Endocytosis in Spinal Muscular Atrophy and Rescue SMA Phenotype". *The American Journal of Human Genetics* 99.3 (2016): 647-665.
45. Pollard TD and Borisy GG. "Cellular motility driven by assembly and disassembly of actin filaments". *Cell* 112.4 (2003): 453-465.
46. Delanote V., *et al.* "versatile modulators of actin organization in (pa-tho) physiological cellular processes". *Acta Pharmacologica Sinica* 26.7 (2005): 769-779.
47. Oprea GE., *et al.* "Plastin 3 is a protective modifier of autosomal recessive spinal muscular atrophy". *Science* 320.5875 (2008): 524-527.
48. Hao T., *et al.* "Survival motor neuron affects plastin 3 protein levels leading to motor defects". *Journal of Neuroscience* 32.15 (2012): 5074-5084.
49. Dubowitz V. "Very severe spinal muscular atrophy (SMA type 0): an expanding clinical phenotype". *European Journal of Paediatric Neurology* 3.2 (1999): 49-51.
50. MacLeod MJ., *et al.* "Prenatal onset spinal muscular atrophy". *European Journal of Paediatric Neurology* 3.2 (1999): 65-72.
51. Sarnat HB and Trevenen CL. "Motor neuron degeneration in a 20-week male fetus: spinal muscular atrophy type 0". *Canadian Journal of Neurological Sciences* 34.2 (2007): 215-220.
52. Wathiyati MS., *et al.* "Combination of SMN2 copy number and NAIP deletion predicts disease severity in spinal muscular atrophy". *Brain and Development* 31.1 (2009): 42-45.
53. Parra J., *et al.* "Ultrasound evaluation of fetal movements in pregnancies at risk for severe spinal muscular atrophy". *Neuromuscular Disorders* 21.2 (2011): 97-101.
54. Munsat TL and Davies KE. "International SMA consortium meeting". *Neuromuscular Disorders* 2 (1992): 423-428.
55. Bertini E., *et al.* "134th ENMC International Workshop: outcome measures and treatment of spinal muscular atrophy. Naarden, The Netherlands". *Neuromuscular Disorders* 15.11 (2005): 802-816.
56. Zerres K., *et al.* "A collaborative study on the natural history of childhood and juvenile onset proximal spinal muscular atrophy (type II and III): 569 patients". *Journal of the Neurological Sciences* 146.1 (1997): 67-72.
57. Rudnik-Schoneborn S., *et al.* "The predictive value of achieved motor milestones assessed in 441 patients with infantile spinal muscular atrophy types II and III". *European Neurology* 45.3 (2001): 174-181.
58. Wirth B., *et al.* "Mildly affected patients with spinal muscular atrophy are partially protected by an increased SMN2 copy number". *Human Genetics* 119.4 (2006): 422-428.
59. Harada Y., *et al.* "Correlation between SMN2 copy number and clinical phenotype of spinal muscular atrophy: three SMN2 copies fail to rescue some patients from disease severity". *Journal of Neuroscience* 249.9 (2004): 1211-1219.
60. Brahe C., *et al.* "Genetic homogeneity between childhood-onset and adult-onset autosomal recessive spinal muscular atrophy". *Lancet* 346.8977 (1995): 741-742.
61. Clermont O., *et al.* "SMN gene deletions in adult-onset spinal muscular atrophy". *Lancet* 346 (1995): 1712-1713.
62. Prior TW., *et al.* "Homozygous SMN1 deletions in unaffected family members and modification of the phenotype by SMN2". *American Journal of Medical Genetics Part A* 130A.3 (2004): 307-310.
63. Burghes AH. "When is a deletion not a deletion? When it is converted?". *American Journal of Human Genetics* 61.1 (1997): 9-15.
64. Hausmanowa-Petrusewicz I., *et al.* "Chronic proximal spinal muscular atrophy of childhood and adolescence: sex influence". *Journal of Medical Genetics* 21 (1984): 447-450.
65. Helmken C., *et al.* "Evidence for modifying pathway in SMA discordant families: reduced SMN level decreases the amount of its interacting partners and Htra2-beta1". *Human Genetics* 114.1 (2003): 11-21.
66. Cusco I., *et al.* "SMN2 copy number predicts acute or chronic spinal muscular atrophy but does not account for intrafamilial variability in siblings". *Journal of Neurology* 253.1 (2006): 21-25.
67. Novelli G., *et al.* "A possible role of NAIP gene deletions in sex-related spinal muscular atrophy phenotype variation". *Neurogenetics* 1.1 (1997): 29-30.

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