



The Effects of $\alpha 7n$ -Acetylcholine, $\beta 2$ -Adrenergic Receptors Agonists, NF- κ B, STAT3 Inhibitors on the Implementation of the Cholinergic Anti-Inflammatory Pathway in Sepsis

Pavel F Zabrodskii*

Saratov Medical University "REAVIZ", Saratov, Russian Federation, Russia

*Corresponding Author: Pavel F Zabrodskii, Saratov Medical University "REAVIZ", Saratov, Russian Federation, Russia.

Received: April 09, 2021

Published: June 04, 2021

© All rights are reserved by **Pavel F Zabrodskii**.

Abstract

Experiments on random-bred albino mice showed that the administration of $\alpha 7n$ -acetylcholine receptors agonist (GTS-21, 15 mg/kg), $\beta 2$ -adrenoreceptor agonist (dexmedetomidine hydrochloride, 25 μ g/kg), NF- κ B inhibitor (BAY 11-7082, 10 mg/kg) and STAT3 inhibitor (S3I-201, 15 mg/kg) to mice 2h before sepsis modeling (i.p, 2.5×10^9 CFUs diurnal culture of *E. coli* O157:H7) caused a significant reduction of mouse mortality at 4 and 24h after *E. coli* injection due to a decrease in the proinflammatory cytokines TNF- α , IL-1 β and IL-6 (implementation of a cholinergic anti-inflammatory pathway). The effects of the preparations used did not differ significantly.

Keywords: Cholinergic Anti-inflammatory Pathway; Sepsis; Proinflammatory Cytokines; $\alpha 7n$ -Acetylcholine Receptor; $\beta 2$ -Adrenoreceptor; NF- κ B Inhibitor; STAT3 Inhibitor

Introduction

Sepsis is a serious public health problem. Worldwide, the incidence of sepsis ranges from 20 to 30 million cases per year, with the frequency of lethality increasing [1-4]. From all deaths associated with diseases and their complications, mortality from sepsis, depending on various factors, ranges from 12 to 60% [5]. The lethality from septic shock, the most severe form of sepsis, continues to be above 50% [6]. We discovered the cholinergic anti-inflammatory mechanism in 1987 [7], named the "cholinergic anti-inflammatory pathway" after studying its implementation at the organismal, systemic, organ, cellular and subcellular levels in 2000 - 2002 [6-10]. We proved the possibility of using cholinomimetics for emergency activation of antimicrobial resistance in sepsis in 1995 [9,10]. The cholinergic anti-inflammatory pathway [3,6,11,12] is realized as a result of activation of acetylcholine m acetylcholine receptors type 1 (m1AChRs) of the brain, modulating the immune regulating function of the vagus nerve; excitation of efferent fibers n. vagus; action of acetylcholine on $\alpha 7n$ -acetylcholine receptors ($\alpha 7n$ AChRs) cells of the monocyte-macrophage system. In the cells of this system, the onset of anti-inflammatory effect is provided by kinase JAK2; signal transducer and activator of transcription 3 (transcription factor) - STAT3; transcription factor NF- κ B (nuclear factor kappa B, NF-kappa B). Activation of

$\alpha 7n$ AChRs of the monocyte-macrophage system by acetylcholine, influencing the function of these biochemical factors, leads to a decrease in the lethality from sepsis due to inhibition of production by the cells of the monocyte-macrophagal system of proinflammatory cytokines TNF- α , IL-1 β , IL-6, B1-HMGB1 protein, macrophage-inflammatory protein- 2 - MIP-2 [3,4,8,11,13]. The similar effect is caused by adrenergic anti-inflammatory mechanism [3] (activation of epinephrine and norepinephrine of $\beta 2$ -adrenoreceptors - $\beta 2$ ARs - monocyte-macrophage system and spleen T-lymphocytes due to excitation of sympathetic ganglia and adrenal medulla by n-acetylcholine receptors) [3,8,14,15].

The comparative study of the effects of $\alpha 7n$ AChRs, $\beta 2$ ARs agonists NF- κ B and STAT3 inhibitors in sepsis is of current interest to develop methods to reduce mortality in this complication [3,4,16-19].

Aim of the Study

The aim of the study was to comparative evaluate the $\alpha 7n$ -acetylcholine receptors (GTS-21) agonist, $\beta 2$ -adrenoreceptors agonist (dexmedetomidine hydrochloride), NF- κ B inhibitor (BAY 11-7082) and STAT3 inhibitor (S3I-201) on the mice mortality after sepsis modeling caused by experimental peritonitis and the content of proinflammatory cytokines TNF- α , IL-1 β , IL-6 in the blood.

Materials and Methods

Experiments were carried out on random-bred albino mice of both sexes weighing 18 - 22g. The control group of mice (control group 1, n = 9) received intraperitoneally 2.0 ml of isotonic sodium chloride solution (saline) 2h after the intraperitoneal administration of 0.5 ml of 0.05% aqueous solution of dimethyl sulfoxide - DMSO (Sigma-Aldrich). The second group of mice (control group 2, n = 160) was injected with 0.5 ml of a 0.05% aqueous solution of DMSO (i.p.). The mice in this group at 2h after administration of this solution received (i.p.) 2.5×10^9 CFUs diurnal culture of *E. coli* O157:H7 in 2.0 ml of saline (sepsis modeling) [1,7,9,20]. The third group of mice was given (i.p., n = 45) the $\alpha 7n$ AChRs agonist GTS-21 [3-(2,4-dimethoxybenzylidene)-anabaseine dihydrochloride] (Sigma-Aldrich) at single dose of 15 mg/kg in 0.5 ml of a 0.05% aqueous solution of DMSO [21]. As the selective agonist of $\beta 2$ ARs (dexmedetomidine hydrochloride - Orion Pharma) was used, which was administered (i.p.) at single dose of 25 μ g/kg [22] in 0.5 ml of a 0.05% aqueous solution of DMSO (group 4, n = 42). The fifth group (n = 38) was administered (i.p.) of NF- κ B inhibitor BAY 11-7082 (Sigma-Aldrich) at single dose of 10 mg/kg in 0.5 ml of a 0.05% aqueous solution of DMSO [19]. The sixth group (n = 40) was administered (i.p.) STAT3 inhibitor (S3I-201 - 2-Hydroxy-4-[[[(4-methylphenyl)sulfonyl]oxy]acetyl]amino]-benzoic acid) (Sigma-Aldrich) at single dose of 15 mg/kg in 0.5 ml of a 0.05% aqueous solution of DMSO [17]. The mice in groups 3 - 6 received at 2h after administration of $\alpha 7n$ AChRs agonist, $\beta 2$ ARs agonist, NF- κ B and STAT3 inhibitors received (i.p.) 2.5×10^9 CFU daily culture of *E. coli* O157:H7 in 2.0 ml saline (sepsis modeling).

The mortality of mice (groups 2 - 6) was recorded at 4 and 24h after the sepsis modelling the concentration of TNF- α , IL1 β and IL-6 were measured in the blood plasma of all groups of mice (groups 1 - 6) using by ELISA (MyBioSource) according to manufacturer's instructions (at 4 and 24h after the sepsis modeling). Determination the concentrations of proinflammatory cytokines used monoclonal antibodies MyBioSource (cat. N - MBS494184, MBS494492, MBS335516 for TNF- α , IL-1 β and IL-6, respectively). Blood for research was taken from the retroorbital venous sinus. The obtained data were processed statistically using Student's t-test. Differences between the parameters were considered reliable at p < 0.05.

Results

The $\alpha 7n$ AChRs agonist (GTS-21), $\beta 2$ ARs agonist (dexmedetomidine hydrochloride), NF- κ B inhibitor (BAY 11-7082) and STAT3 inhibitor (S3I-201) caused a decrease in the mortality of mice 4h

after the administration of the daily culture of *E. coli* (sepsis modeling) compared to the control group 2 (sepsis) by 2.5 times (by 30.0%), by 1.9 times (by 23.8%), by 1.7 times (by 21.1%) and by 1.5 times (by 17.5%) respectively (p < 0.05). The $\alpha 7n$ AChRs agonist, $\beta 2$ ARs agonist, NF- κ B and STAT3 inhibitors caused a decrease in the mortality of mice 24h after the sepsis modeling compared to the control group 2 (sepsis) by 1.4 times (by 24.7%), by 1.5 times (by 27.7%), by 1.7 times (by 32.5%) and by 1.3 times (by 17.5%) respectively (p < 0.05). The reduction in mortality under the different preparations (groups 3 - 5) was not significantly different (p > 0.05) (Table 1).

Table 1: Effects of $\alpha 7n$ -acetylcholine receptors agonist (GTS-21, 15 mg/kg), $\beta 2$ -adrenoreceptor agonist (dexmedetomidine hydrochloride, 25 μ g/kg), NF- κ B inhibitor (BAY 11-7082, 10 mg/kg), and STAT3 inhibitor (S3I-201, 15 mg/kg) on the mice mortality after sepsis modeling, % (M \pm m).

Series of experiments	Term study of mortality after administration of <i>E. coli</i> , h	
	4	24
Sepsis (control group 2, n = 160)	50.0 \pm 4,0	82,5 \pm 3,0
$\alpha 7n$ AChRs agonist (GTS-21) + sepsis (group 3; n = 45)	20,0 \pm 6,0*	57,8 \pm 7,4*
$\beta 2$ ARs agonist hexaprenaline sulfate + sepsis (group 4, n = 42)	26.2 \pm 6.8*	54,8 \pm 7.8*
NF- κ B inhibitor BAY 11-7082 + sepsis (group 5, n = 38)	28,9 \pm 7,5*	50,0 \pm 8,2*
STAT3 inhibitor - S3I-201+ sepsis (group 6, n = 40)	32.5 \pm 7.5*	65.0 \pm 7.6*

* -p < 0,05 as compared to control (group 2).

The concentration of cytokines TNF- α , IL-1 β and IL-6 after the sepsis modeling (control group 2) in the blood of mice after 4h compared with the control group 1 (intact animals), increased respectively by 18.5; 18.1 and 50.5 times (p < 0.05) and after 24h, the content of these cytokines compared to their level after 4h decreased, respectively, by 13.2; 4.8 and 8.4 times (p < 0.05). The content of IL-1 β and IL-6 after 24h remained higher than in group 1 by 1.6 times (p > 0.05) and 4.9 times (p < 0.05), respectively, and the concentration of TNF- α in groups 1 and 2 did not differ significantly (Table 2).

The $\alpha 7n$ AChRs agonist (GTS-21) at 4h after sepsis modeling reduced the blood levels of TNF- α , IL-1 β and IL-6 (group 3) compared to the control group 2, respectively, by 3.6; 3.8 and 9.1 times

Table 2: Effects of $\alpha 7n$ -acetylcholine receptors agonist (GTS-21, 15 mg/kg), $\beta 2$ -adrenoreceptor agonist (dexmedetomidine hydrochloride, 25 μ g/kg), NF- κ B inhibitor (BAY 11-7082, 10 mg/kg) and STAT3 inhibitor (S3I-201, 15 mg/kg) on the concentration of proinflammatory cytokines in blood of mice after sepsis modeling, pg/ml ($M \pm m$; n=8-9)

Series of experiments	TNF- α		IL-1 β		IL-6	
	4	24	4	24	4	24
Control group 1	42 \pm 5	38 \pm 6	35 \pm 6	27 \pm 7	45 \pm 7	25 \pm 5
Sepsis (Control group 2)	780 \pm 90 ^a	59 \pm 9 ^c	635 \pm 73 ^a	132 \pm 25 ^{ac}	2273 \pm 263 ^a	271 \pm 38 ^{ac}
$\alpha 7n$ AChRs agonist (GTS-21) + sepsis (group 3)	151 \pm 22 ^{ab}	49 \pm 9 ^c	167 \pm 30 ^{ab}	54 \pm 8 ^{abc}	250 \pm 33 ^{ab}	82 \pm 14 ^{abc}
$\beta 2$ ARs agonist (dexmedetomidine hydrochloride) + sepsis (group 4)	169 \pm 28 ^{ab}	52 \pm 8 ^c	184 \pm 32 ^{ab}	64 \pm 10 ^{abc}	295 \pm 35 ^{ab}	105 \pm 14 ^{abc}
NF- κ B inhibitor (BAY 11-7082) + sepsis (group 5)	129 \pm 22 ^{ab}	47 \pm 7 ^c	120 \pm 24 ^{ab}	52 \pm 8 ^{abc}	220 \pm 29 ^{ab}	75 \pm 9 ^{abc}
STAT3 inhibitor (S3I-201) + sepsis (group 6)	137 \pm 30 ^{ab}	43 \pm 8 ^c	155 \pm 36 ^{ab}	70 \pm 11 ^{abc}	312 \pm 38 ^{ab}	94 \pm 10 ^{abc}

Note. 4 and 24 - time after sepsis modeling, h; in parentheses - number of animals; ^a - p < 0.05 compared to control (group 1); ^b - p < 0.05 compared to the corresponding parameter of control group 2 (sepsis modeling); ^c - p < 0.05 compared with parameter after 4 h.

(p < 0.05). The content of these cytokines after 24h compared with their level after 4h decreased, respectively, by 3.1; 2.4 and 3.3 times (p < 0.05). The concentrations of IL-1 β and IL-6 after 24h statistically significantly (p < 0.05) exceeded those of the control group 1 by 2.0 and 3.3 times (p < 0.05), respectively and compared to the parameters of group 2, the content IL-1 β and IL-6 were reduced by 2.4 and 3.3 times respectively (p < 0.05).

The $\beta 2$ ARs agonist (dexmedetomidine hydrochloride) at 4h after sepsis modeling decreased the blood concentrations of TNF- α , IL-1 β and IL-6 (group 4) compared to the control group 2 by 4.6; 3.5 and 7.7 times respectively (p < 0.05). The blood levels of TNF- α , IL-1 β and IL-6 after 24h compared with their level after 4h decreased, respectively, by 3.3; 2.9 and 2.8 times (p < 0.05). The concentrations of IL-1 β and IL-6 after 24h were significantly higher (p < 0.05) than those of the control group 1 by 2.4 and 4.2 times (p < 0.05), respectively, and compared to the parameters of group 2, the blood concentrations of IL-1 β and IL-6 were decreased by 2.6 times (p < 0.05).

The NF- κ B inhibitor (BAY 11-7082) at 4h after sepsis modeling reduced the blood levels of TNF- α , IL-1 β and IL-6 (group 5) compared to the control group 2, respectively, by 6.1; 5.3 and 10.3 times (p < 0.05). The content of these cytokines after 24h compared with their level after 4h decreased, respectively, by 2.7; 2.3 and 2.9 times (p < 0.05). The concentrations of IL-1 β and IL-6 after 24h statistically significantly (p < 0.05) exceeded those of the control group 1 by 1.9 and 3.0 times (p < 0.05), respectively, and compared to the parameters of group 2, the content IL-1 β and IL-6 were reduced by 2.5 and 3.6 times respectively (p < 0.05).

The STAT3 inhibitor (S3I-201) at 4h after sepsis modeling de-

creased the blood concentrations of TNF- α , IL-1 β and IL-6 (group 6) compared to the control group 2 by 5.7; 1.9 and 7.3 times respectively (p < 0.05). The blood levels of TNF- α , IL-1 β and IL-6 after 24h compared with their level after 4h decreased, respectively, by 3.2; 2.2 and 3.3 times (p < 0.05). The concentrations of IL-1 β and IL-6 after 24h were significantly higher (p < 0.05) than those of the control group 1 by 2.4 and 4.2 times (p < 0.05), respectively, and compared to the parameters of group 2, the content IL-1 β and IL-6 were reduced, respectively, by 1.9 and 2.9 times (p < 0.05).

The value of TNF- α was not significantly different from the levels in groups 1 - 6 at 24h after the sepsis modeling. The blood concentrations of proinflammatory cytokines TNF- α , IL-1 β and IL-6 at 4 and 24h after sepsis modeling with the $\alpha 7n$ -acetylcholine receptors agonist (GTS-21), $\beta 2$ -adrenoreceptor agonist (dexmedetomidine hydrochloride), NF- κ B inhibitor (BAY 11-7082) and STAT3 inhibitor (S3I-201) administration (groups 3-6) were not significantly different from those of control group 2 (sepsis modeling without the use of preparations).

Discussion

The $\alpha 7n$ AChRs agonist (GTS-21) due to the implementation of the cholinergic anti-inflammatory pathway [4,11,23] leads to a reduction in sepsis mortality [7,9,10,24] as a result of decrease of the production of proinflammatory cytokines by cells of the monocyte-macrophage system [3,4,13,24-26]. The $\beta 2$ ARs agonist (hexaprenaline sulfate) caused a similar effect. When $\beta 2$ ARs and $\alpha 7n$ AChRs agonists were used, there was no significant difference in mouse mortality between the parameters in these groups (3 and 4) after sepsis modeling. The adrenergic mechanism (the action of the $\beta 2$ ARs agonist) is an important component in the implementa-

tion of the cholinergic anti-inflammatory pathway [3]. Monocytes and macrophages are known to have β ARs, and their activation, results in reduced synthesis of proinflammatory cytokines due to impaired Ca^{2+} exchange and probably inhibition of the nuclear transcription factor NF- κ B [5,14,27].

The reduction in mortality in mice after sepsis modeling under the influence of NF- κ B inhibitor (BAY 11-7082) and STAT3 inhibitor (S3I-201) is due to a decrease in proinflammatory cytokine production (TNF- α , protein B1 - HMGB1, macrophage inflammatory protein-2 - MIP-2, interleukins - IL-1 β , IL-6) by cells of the monocyte-macrophage system [4,11,24,28,29]. The transcription factor NF- κ B modulates the synthesis of proinflammatory cytokines involved in the development of sepsis. Signalling pathways initiated by Toll-like receptors (TLR2 and TLR4) to which bacterial products, particularly *E. coli* lipopolysaccharide, bind lead to increased transcription of genes responsible for the expression of cytokines, chemokines, adhesion molecules, apoptotic factors and other mediators of sepsis-related inflammatory response [19].

The absence of statistically significant differences in mortality and blood concentrations of TNF- α , IL-1 β , IL-6 in sepsis models under the action of $\alpha 7$ nAChRs agonist (GTS-21), $\beta 2$ ARs agonist (dexmedetomidine hydrochloride), NF- κ B inhibitor (BAY 11-7082) and STAT3 inhibitor (S3I-201) is due to the fact that these drugs were used in equivalent (equally therapeutic) doses [3,4,17,19,21,22,24,30]. Their effect is ultimately realized by reducing the production of proinflammatory cytokines [4,11,28,29].

Conclusion

The administration of $\alpha 7$ n-acetylcholine receptors agonist (GTS-21, 15 mg/kg), $\beta 2$ -adrenoreceptors agonist (dexmedetomidine hydrochloride, 25 μ g/kg), NF- κ B inhibitor (BAY 11-7082, 10 mg/kg) and STAT3 inhibitor (S3I-201, 15 mg/kg) to mice 2h before sepsis modeling (i.p. *E. coli* O157:H7) caused a significant reduction of mouse mortality at 4 and 24h after *E. coli* injection due to a decrease in the proinflammatory cytokines TNF- α , IL-1 β and IL-6 (implementation of the cholinergic anti-inflammatory pathway). The effects of the preparations used did not differ significantly.

Bibliography

1. Martin GS. "Sepsis, severe sepsis and septic shock: changes in incidence, pathogens and outcomes". *Expert Review of Anti-infective Therapy* 10.6 (2012): 701-706.

2. Byrne L and Van Haren F. "Fluid resuscitation in human sepsis: Time to rewrite history?". *Annals of Intensive Care* 7.1 (2017): 4.
3. Zabrodskii PF, et al. "Role of $\beta 2$ -adrenoreceptors in adrenergic anti-inflammatory mechanism in sepsis". *Bulletin of Experimental Biology and Medicine* 162.12 (2016): 718-721.
4. Zabrodskii PF, et al. "Combined effect of NF- κ B inhibitor and $\beta 2$ -Adrenoreceptor agonist on mouse mortality and blood concentration of proinflammatory cytokines in sepsis". *Bulletin of Experimental Biology and Medicine* 165.4 (2018): 445-448.
5. Lin JN, et al. "Risk factors for mortality of bacteremic patients in the emergency department". *Academic Emergency Medicine* 16 (2009): 749-755.
6. Bernik TR, et al. "Pharmacological stimulation of the cholinergic anti-inflammatory pathway". *Journal of Experimental Medicine* 195.6 (2002): 781-788.
7. Zabrodskii PF. "Effect of armin on nonspecific resistance factors of the body and on the primary humoral immune response". *Farmakologiya i Toksikologiya* 50.1 (1987): 57-60.
8. Bonaz BL and Bernstein CN. "Brain-gut interactions in inflammatory bowel disease". *Gastroenterology* 144.1 (2013): 36-49.
9. Zabrodskii PF. "Variation in anti-infectious nonspecific resistance of the organism caused by cholinergic stimulation". *Bulletin of Experimental Biology and Medicine* 120.2 (1995): 809-811.
10. Zabrodskii PF. "Change in the non-specific anti-infection resistance of the body exposed to cholinergic stimulation". *Bulletin of Experimental Biology and Medicine* 120.8 (1995): 164-166.
11. Borovikova LV, et al. "Vagus nerve stimulation attenuates the systemic inflammatory response to endotoxin". *Nature* 405.6785 (2000): 458-462.
12. Kessler W, et al. "The vagal nerve as a link between the nervous and immune system in the instance of polymicrobial sepsis". *Langenbeck's Archives of Surgery* 391.2 (2006): 83-87.
13. Kox M and Pickkers P. "Modulation of the innate immune response through the vagus nerve". *Nephron* 131.2 (2015): 79-84.

14. Scanzano A and Cosentino M. "Adrenergic regulation of innate immunity: a review". *Frontiers in Pharmacology* 23.6 (2015): 171.
15. Zetterlund A., et al. "Effects of beta2-agonist and bedesonide on interleukin-lbeta and leukotriene B4 secretion: studies of human monocytes and alveolar macrophages". *Journal of Asthma* 35.7 (1998): 565-573.
16. Abraham E. "Nuclear factor-kappaB and its role in sepsis-associated organ failure". *Journal of Infectious Diseases* 187 (2003): 364-369.
17. Park JW., et al. "Inhibition of STAT3 activity delays obesity-induced thyroid carcinogenesis in a mouse model". *Endocrine-Related Cancer* 23.1 (2016): 53-63.
18. Wang Z., et al. "The STAT3 inhibitor S3I-201 suppresses fibrogenesis and angiogenesis in liver fibrosis". *Laboratory Investigation* 98.12 (2018): 1600-1613.
19. Zhang H., et al. "Hydrogen sulfide acts as an inflammatory mediator in cecal ligation and puncture-induced sepsis in mice by upregulating the production of cytokines and chemokines via NF-kappaB". *American Journal of Physiology - Lung Cellular and Molecular Physiology* 292.4 (2007): 960-971.
20. Song DJ., et al. "Effect of lentiviral vector encoding on triggering receptor expressed on myeloid cells 1 on expression of inflammatory cytokine in septic mice infected by *Bacteroides fragilis*". *Zhonghua Shao Shang Za Zhi* 25.1 (2009): 36-41.
21. Norman GJ., et al. "Cardiopulmonary arrest and resuscitation disrupts cholinergic anti-inflammatory processes: a role for cholinergic $\alpha 7$ nicotinic receptors". *Journal of Neuroscience* 31.9 (2011): 3446-3452.
22. Ning Q., et al. "Neurodegenerative changes and neuroapoptosis induced by systemic lipopolysaccharide administration are reversed by dexmedetomidine treatment in mice". *Neurology Research* 39.4 (2017): 357-366.
23. Eftekhari G., et al. "Activation of central muscarinic receptor type 1 prevents development of endotoxin tolerance in rat liver". *European Journal of Pharmacology* 740 (2014): 436-441.
24. PF Zabrodskii. "Immunotoxicology of organophosphorus compounds". *Saratov* (2016): 289.
25. Martelli D., et al. "The cholinergic anti-inflammatory pathway: a critical review". *Autonomic Neuroscience* 182 (2014): 65-69.
26. Zabrodskii PF, et al. "Effect of $\alpha 7$ n-Acetylcholine Receptor Activation and Antibodies to TNF- α on Mortality of Mice and Concentration of Proinflammatory Cytokines During Early Stage of Sepsis". *Bulletin of Experimental Biology and Medicine* 159.6 (2015): 740-742.
27. Tan KS., et al. "Beta2 adrenergic receptor activation stimulates pro-inflammatory cytokine production in macrophages via PKA- and NF-kappaB-independent mechanisms". *Cell Signaling* 19.2 (2007): 251-260.
28. Gallowitsch-Puerta M., et al. "Neuro-immune interactions via the cholinergic anti-inflammatory pathway". *Life Science* 80.24-25 (2007): 2325-2329.
29. Pavlov VA., et al. "The cholinergic anti-inflammatory pathway: a missing link in neuroimmunomodulation". *Molecular Medicine* 9.5-8 (2003): 125-134.
30. Zabrodskii PF, et al. "Combined Effects of M1 Muscarinic Acetylcholine Receptor Agonist TBPB and $\alpha 7$ n-Acetylcholine Receptor Activator GTS-21 on Mouse Mortality and Blood Concentration of Proinflammatory Cytokines in Sepsis". *Bulletin of Experimental Biology and Medicine* 162.6 (2017): 750-753.

Volume 5 Issue 7 July 2021

© All rights are reserved by Pavel F Zabrodskii.