

Endoplasmic Reticulum Aminopeptidase-1 (ERAP-1) Mutations in People with Familial Mediterranean Fever (FMF) Gene Mutations Without Disease

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Abstract

Background and Study Aim: To date, especially alterations of genes on exon 10 have been considered in Mediterranean fever (MEFV), but it is not clear whether all these alterations are disease-causing mutations.

We studied ERAP-1 gene mutations in people with FMF gene mutations without FMF phenotype. We compared our results with our previous study which we studied ERAP-1 gene mutations in patients with FMF and with ulcerative colitis (which has been shown to have a relation with ERAP-1 gene mutations before).

Patients and Methods: This is a retrospective study. We studied ERAP-1 gene mutation in blood samples (had been kept in -70°C before for different studies) from 51 people with FMF gene mutations without FMF disease. Some of these people also were followed for about 5 years whether FMF comes out.

Results: There were 10 cases without ERAP-1 mutations at Exon-3 and 6 cases without ERAP-1 mutations at Exon-10. Any ERAP-1 mutations were not obtained in three cases both for Exon-3 and 10.

There were 41 ERAP-1 gene mutations at Exon-3 and 48 at Exon-10. Exon-3 mutations were usually Codon 127 mutation for 35 patients whether it was single mutation or with multiple mutations. It was totally different our ERAP-1 mutation in patients with FMF for Exon-3. Because in our previous study, if there were ERAP-1 gene mutations at exon-3, [c.380G>C p.(Arg127Pro)] mutation always existed in patients with FMF.

Conclusion: As a conclusion, ERAP-1 may be the second needed genetic mutation to produce the disease, 2) Certain ERAP-1 genetic mutation may produce FMF together with certain pyrin gene mutation.

Keywords: Familial Mediterranean Fever; ERAP-1; Inflammation; M694

Introduction

Familial Mediterranean Fever (FMF) is an autosomal recessive and accepted as a monogenic disorder that frequently affects Turkish, North Africans, Jewish, Arabic, and Armenian populations [1]. It is characterized by episodic abdominal pain, arthritis, pericarditis, pleurisy, fever and skin rash. However, effort of explaining the pathophysiology of the disease provide to find the genetical back-

ground of the FMF. Finding the pyrin mutation, made a new line of vision to not only to FMF but all the autoinflammatory diseases [1-13].

However, we presumed that single mutations in pyrin gene is not enough to produce FMF phenotype. Because we believe that to accept this disease as monogenic with its heterogeneous clinical aspects may be contradictory. There may be some additional

mutations for the manifestations of FMF disease out of *pyrin* gene mutations. In our previous study we almost had showed that there are more *ERAP-1* mutations (especially in exon10) in patients with FMF in accordance with patients in ulcerative colitis group which had been shown the relation before [1]. In the light of these results, we concluded that there may be a strong susceptibility with *ERAP-1* gene mutations for occurring clinics of FMF. All patients with FMF had FMF mutations (mostly M694) in our previous study. Therefore, our FMF population was quite homogeneous.

The *ERAP1* gene provides instructions for making a protein called endoplasmic reticulum aminopeptidase. Basically, it regulates aminopeptidase which is an enzyme that cleaves other proteins into small peptides to provide the recognizing the antigenic determinants. If the immune system recognizes these peptides as foreign (such as viral or bacterial peptides) it responds by triggering the infected cell to self-destruction [12-17].

Therefore, in this current study we studied *ERAP-1* gene mutations in people with *FMF* gene mutations but without FMF disease (without phenotype). We tried to verify that manifest FMF disease needs additional mutation(s) by showing people with *MEFV* gene mutations but without disease. We also compared our results with our previous study which we studied *ERAP-1* gene mutations (which is possible additive effect on *MEFV* gene mutations) in patients with FMF and with ulcerative colitis (which has been shown to have a relation with *ERAP-1* gene mutations before).

Materials and Methods

This is a kind of retrospective study. We studied *ERAP-1* gene mutation in blood samples (which had been kept in -70°C before for some previous studies) from 51 people with *FMF* gene mutations without FMF disease. They invite again to our center, if they had any attacks of the disease during the out of control of us. If the cases with *MEFV* gene mutations had any FMF attacks during this uncontrolled period was not included to the normal phenotype group. Some of these people also had been followed for about 5 years but any FMF clinics has not been happened.

The mean age of the cases was 38.90 ± 7.09 (M/F: 15/36) years.

Helsinki and local approval was received from the Ethics Committee of Training Hospital of Maltepe Medical Faculty Maltepe -İstanbul. Informed consent was obtained from all patients.

Detection of serum *ERAP-1* gen mutations

Deposited serums in Eppendorf tubes at -70°C were used for *ERAP-1* mutations. Analysis of *ERAP-1* gene mutations at exon-3 and exon-10 were done with PCR-DNA folders. Primer pairs were used at 0.6 pico mole/mL. The other PCR components were 10 mM Tris-HCl (25°C pH: 8.8), 50 mM KCl, 0.2 mM deoksi-nukleotit-3-phosphate [dATG, dGTP, dCTP, dTTP (Fermentas, Lithuania)] and 15 mM MgCl_2 . *ERAP-1* gene primers at exon-3: 5'AGT TCA ACA GCA AAG GGA ATT3', 5'TTT TGC TTT TGT ACA TTT G3' *ERAP-1* gene primers at exon-10: 5'CTC CTC AGA GGG ATT AAC ATA, 5'TTA ACA GTG TTC CTG CAG TTG CG3'. After denaturation and hybridization (Applied Biosystems, USA) of PCR-products, amplification of right gene region was checked by agarose gel.

After purification of samples (Bio Basic, Canada), a new PCR-cycling was done and purified again. At the end of this process, samples were loaded to the DNA folders analyzer (Applied Biosystems 3130xl, USA). Obtained results were analyzed by SeqScape® Software v3.0.

Results were given as both first cDNA and and second protein changes.

Statistical analysis

Statistical analysis was done by SSPS statistical software (SPSS for windows 10.0, Inc., Chicago, IL, USA). Groups were compared with Marginal Homogeneity. Statistical significance was defined as $p < 0.05$.

Results

The results are shown in figure 1. There were 34 cases with single mutation (18 cases with M694V mutation, 5 cases with E148Q, 4 cases with M680I, 4 cases with V726A, 3 cases with A744S) and 17 cases with multiple mutations (5 cases with M694V and E148Q, 1 case with M694V and 680I, 4 cases with M694V and V726A, 2 cases with M680I and V726A, 4 cases with E148Q and P369S, 1 cases with M680I and E148Q).

M694V.....	: 18	} Single mutation: 34
E148Q.....	: 5	
M680I.....	: 4	
V726A.....	: 4	
A744S.....	: 3	} Multiple mutations: 17
M694V+E148Q.....	: 5	
M694V+680I.....	: 1	
M694V+V726A.....	: 4	
M680I+V726A.....	: 2	
E148Q+P369S.....	: 4	
M680I+E148Q.....	: 1	
+		
TOTAL	: 51 cases	

Figure 1: The distribution of FMF gene mutations.

There were 11 cases without *ERAP-1* mutations at Exon-3, 6 cases without *ERAP-1* mutations at Exon-10. *ERAP-1* mutations were not obtained in three cases both for Exon-3 and 10 (Table 1).

There were 40 cases with *ERAP-1* gene mutations at Exon-3 and 45 cases with *ERAP-1* gene mutations at Exon-10. Exon-3 mutations were usually c.380 G>C p.(Arg127Pro) mutation for 35 patients whether it was single mutation or with multiple mutations (33 single mutations and 2 multiple mutations) (Table 1). It was totally very similar with our previous study showing *ERAP-1* mutation in patients with FMF for Exon-3 (Table 2).

There were 45 single *ERAP-1* gene mutations at exon-10. [c.1359T>C p.(Ser127Pro)] was the only mutation type for all cases whether it was heterozygote or homozygote. It was also totally different for our previous study in patients with FMF who had single mutation [c.1359T>C p.(Ser453Pro)] in 10 patients and [c.1359T>C

p.(Ser127Pro)] in three patients FMF showed [c.380G>C p.(Arg127Pro)]single gene mutation at exon-10 (Table 2).

Dominant mutation for *ERAP-1* gene [c.380G>C p.(Arg127 Pro)] at Exon-3 in patient group was significantly similar ($p < 0.005$) with non-patient group in contradistinction to being at Exon-10. Dominant mutation at Exon-10 was [c.380G>C p.(Ser 453 Ser)] in patient group. This mutation was significantly different between patient group and non-patient group ($p > 0.7$) at Exon-10.

Discussion and Conclusion

To show the genetic background of FME, it is very important to understand main mechanism of all this kind of auto-inflammatory disease. In previous studies, it was studied to show correlation of one kind of mutation with FMF. Most of the studies had shown that different kinds of Exon-10 mutations were significantly high in different ethnic groups. However, these findings could not be tested.

	No	MEFV Gene Mutation	ERAP-1 Mutation					
			Exon-3			Exon-10		
			cDNA sequence	Protein sequence	Zygoty	cDNA sequence	Protein sequence	Zygoty
Cases with FMF gene mutations without FMF disease	1	V726A	c.420 G>A	p.(Arg127 Pro)	Homozygote	c.1359T>C	p.(Ser127 Ser)	Homozygote
	2	M694V/ V726A	c.36 G>C	p.(Ile/I Thr)	Homozygote	c.1359T>C	p.(Ser127 Ser)	Heterozygote
	3	M680I/ V726A	Normal			c.1359T>C	p.(Ser127 Ser)	Homozygote
	4	M694V	c.380G>C	c.1359T>C	p.(Ser127 Ser)	c.1359T>C	p.(Ser127 Ser)	Heterozygote
	5	M694V/ V726A	c.36 C>T	c.1359T>C	p.(Ser127 Ser)	c.1359T>C	p.(Ser127 Ser)	Heterozygote
	6	M694V	c.380G>C	p.(Arg127 Pro)	Homozygote	c.1359T>C	p.(Ser127 Ser)	Homozygote
	7	M694V/E148Q	c.380G>C	p.(Arg127 Pro)	Heterozygote	c.1359T>C	p.(Ser127 Ser)	Homozygote
	8	M694V/ V726A	Normal			c.1359T>C	p.(Ser127 Ser)	Heterozygote
	9	V726A/V726A	c.184 A > A	c.1359T>C	p.(Ser127 Ser)	c.1359T>C	p.(Ser127 Ser)	Heterozygote
			c.380G>C	c.1359T>C	p.(Ser127 Ser)			
	10	E148Q/M694V	c.380G>C	c.1359T>C	p.(Ser127 Ser)	c.1359T>C	p.(Ser127 Ser)	Heterozygote
	11	V726A	Normal			Normal		
	12	E148Q	Normal			Normal		
	13	M694V/M694V	c.380 G>C	p.(Arg127 Pro)	Homozygote	c.1359T>C	p.(Ser127 Ser)	Homozygote
	14	E148Q/M694V	c.380 G>C	p.(Arg127 Pro)	Homozygote	Normal		
	15	E148Q	c.420 G>A	p.(Arg127 Pro)	Homozygote	c.1359T>C	p.(Ser127 Ser)	Homozygote
	16	A744	Normal			Normal		
	17	E148Q/M694V	c.380 G>C	p.(Arg127 Pro)	Heterozygote	c.1359T>C	p.(Ser127 Ser)	Homozygote
	18	M694V	c.380 G>C	p.(Arg127 Pro)	Heterozygote	c.1359T>C	p.(Ser127 Ser)	Homozygote
	19	M694V/M694V	c.380 G>C	p.(Arg127 Pro)	Heterozygote	Normal		
	20	M680I/V726A	c.380 G>C	p.(Arg127 Pro)	Homozygote	c.1359T>C	p.(Ser127 Ser)	Homozygote
			c.420 G>A	p.(Arg127 Pro)	Homozygote			
	21	M680I/M680I	c.380 G>C	p.(Arg127 Pro)	Homozygote	Normal		
			c.130 C>T	p.(Tyr 57Tyr)	Homozygote			
	22	V726/M694V	c.380 G>C	p.(Arg127 Pro)	Homozygote	c.1359T>C	p.(Ser127 Ser)	Heterozygote
	23	M694V	c.380 G>C	p.(Arg127 Pro)	Homozygote	c.1359T>C	p.(Ser127 Ser)	Homozygote
	24	M694V/M694V	c.380 G>C	p.(Arg127 Pro)	Homozygote	c.1359T>C	p.(Ser127 Ser)	Heterozygote
	25	M694V	c.380G>A	p.(Arg127 Pro)	Heterozygote	c.1359T>C	p.(Ser127 Ser)	Homozygote
	26	V726A	c.380 G>C	p.(Arg127 Pro)	Homozygote	c.1359T>C	p.(Ser127 Ser)	Heterozygote
	27	M694V	c.380 G>C	p.(Arg127 Pro)	Homozygote	c.1359T>C	p.(Ser127 Ser)	Homozygote
28	E148Q	c.380 G>C	p.(Arg127 Pro)	Homozygote	c.1359T>C	p.(Ser127 Ser)	Heterozygote	
29	E148Q	c.380 G>C	p.(Arg127 Pro)	Homozygote	c.1359T>C	p.(Ser127 Ser)	Homozygote	
30	M694V	c.380 G>C	p.(Arg127 Pro)	Homozygote	c.1359T>C	p.(Ser127 Ser)	Heterozygote	

31	M694V	c.380 G>C	p.(Arg127 Pro)	Homozygote	c.1359T>C	p.(Ser127 Ser)	Homozygote
32	M680I	Normal			c.1359T>C	p.(Ser127 Ser)	Heterozygote
33	A744S	c.391C>T	c.1359T>C	p.(Ser127 Ser)	c.1359T>C	p.(Ser127 Ser)	Heterozygote
34	M694V	c.380 G>C	c.1359T>C	p.(Ser127 Ser)	c.1359T>C	p.(Ser127 Ser)	Heterozygote
35	M694V/M694V	Normal			c.1359T>C	p.(Ser127 Ser)	Heterozygote
36	E148Q/M694V	c.380 G>C	p.(Arg127 Pro)	Homozygote	c.1359T>C	p.(Ser127 Ser)	Homozygote
37	E148Q/M680I	c.380 G>C	p.(Arg127 Pro)	Homozygote	c.1359T>C	p.(Ser127 Ser)	Homozygote
38	E148Q	c.380 G>C	p.(Arg127 Pro)	Homozygote	c.1359T>C	p.(Ser127 Ser)	Heterozygote
39	E148Q/P369S	c.380 G>C	p.(Arg127 Pro)	Homozygote	c.1359T>C	p.(Ser127 Ser)	Heterozygote
40	A744S	c.380 G>C	p.(Arg127 Pro)	Homozygote	c.1359T>C	p.(Ser127 Ser)	Homozygote
41	E148Q/P369S	c.380 G>C	p.(Arg127 Pro)	Homozygote	c.1359T>C	p.(Ser127 Ser)	Homozygote
42	M694V/M694V	Normal			c.1359T>C	p.(Ser127 Ser)	Heterozygote
43	E148Q/P369S	Normal			c.1359T>C	p.(Ser127 Ser)	Heterozygote
44	M694V/M694V	c.380 G>C	p.(Arg127 Pro)	Homozygote	c.1359T>C	p.(Ser127 Ser)	Homozygote
45	M680I/M680I	c.380 G>C	p.(Arg127 Pro)	Homozygote	c.1359T>C	p.(Ser127 Ser)	Heterozygote
46	M694V/M694V	c.380 G>C	p.(Arg127 Pro)	Homozygote	c.1359T>C	p.(Ser127 Ser)	Heterozygote
47	M694V	Normal			c.1359T>C	p.(Ser127 Ser)	Heterozygote
48	M680I/M694V	c.380 G>C	p.(Arg127 Pro)	Heterozygote	c.1359T>C	p.(Ser127 Ser)	Heterozygote
49	E148Q/E148Q/P369S	Normal			c.1359T>C	p.(Ser127 Ser)	Heterozygote
50	M680I	Heterozygote	c.1359T>C	p.(Ser127 Ser)	c.1359T>C	p.(Ser127 Ser)	Heterozygote
51	M694V	Heterozygote	Heterozygote	Heterozygote	c.1359T>C	p.(Ser127 Ser)	Homozygote

Table 1: Whole documents of the study for patients with FMF mutation without FMF disease.

	Patient Number	EXON-3			Exon-10		
		cDNA sequence	Protein sequence	Zygotity	cDNA sequence	Protein sequence	Zygotity
Familial Mediterranean Fever	1	c.380G>C	p.(Arg127 Pro)	Heterozygote	c.1359T>C	p.(Ser127 Ser)	Heterozygote
	2	c.380G>C c.35C>T	p.(Arg127 Pro) p.(Thr121Ile Pro)	Heterozygote Heterozygote	Normal		
	3	c.380G>C c.35C>T	p.(Arg127 Pro) p.(Thr121Ile Pro)	Homozygote Heterozygote	c.1359T>C	p.(Ser127 Ser)	Heterozygote
	4	Normal			c.1359T>C	p.(Ser127 Ser)	Heterozygote
	5	c.380G>C	p.(Arg127 Pro)	Homozygote	c.1359T>C	p.(Ser453 Pro)	Homozygote
	6	c.380G>C	p.(Arg127 Pro)	Homozygote	c.1359T>C	p.(Ser453 Pro)	Homozygote
	7	c.380G>C	p.(Arg127 Pro)	Heterozygote	c.1359T>C	p.(Ser453 Pro)	Heterozygote

	8	Normal			c.1359T>C	p.(Ser453 Pro)	Heterozygote
	9	Normal			c.1359T>C	p.(Ser453 Pro)	Heterozygote
	10	Normal			c.1359T>C	p.(Ser453 Pro)	Heterozygote
	11	c.380G>C c.35C>T c.171C>T	p.(Arg127 Pro) p.(Thr127 Pro) p.(Tyr57Pro)	Homozygote Heterozygote Heterozygote	c.1359T>C	p.(Ser453 Pro)	Heterozygote
	12	c.380G>C	p.(Arg127 Pro)	Heterozygote	c.1359T>C	p.(Ser453Ser)	Homozygote
	13	c.380G>C	p.(Arg127 Pro)	Homozygote	c.1359T>C	p.(Ser453Ser)	Homozygote
	14	c.380G>C	p.Arg127 Pro	Heterozygote	c.1359T>C	p.(Ser453Ser)	Homozygote
	15	c.380G>C	p.Arg127 Pro	Heterozygote	c.380G>C	p.Arg127 Pro	Heterozygote

Table 2: Results of *ERAP-1* gene mutations in patients with FMF*.

*All of the patient.

Because there is no study done on cases with *FMF* gene mutation without FMF. On the other hand, FMF can be a polygenic disease according to our thinking of origin.

In this study and in our previous study, for the first time, we showed that if a definite *ERAP-1* and classical *pyrin* gene mutations are together then the possibility of FMF phenotype increases.

In our previous study [c.1359T>C p.(Ser453Pro)] mutation at Exon-10 of *ERAP-1* was more common in patient group with FMF. However, in our current study [c.1359T>C p.(Ser127Ser)] is more common in cases with FMF mutations without disease at Exon-10 for *ERAP-1*. It is [c.380G>C p.(Arg127 Pro)] for cases with FMF and [p.Arg127 Pro (c.380G>C)] for cases without disease at Exon-3. Difference between two group (with cases FMF and without FMF) for *ERAP-1* mutation is more definite at Exon-10 than Exon-3.

We strongly believe that these results are very important. Because this study may show that 1-) *ERAP-1* may be the second needed genetic mutation to produce the disease, 2-) Certain *ERAP-1* genetic mutation may produce FMF together with certain *pyrin* gene mutation.

FMF is a very heterogeneous disease which was shown to have *pyrin* gene mutation before. So, there may be also some other mu-

tations together in the same patients. *ERAP-1* gene controls a very important enzyme which helps to recognize the proteins. This step is a key step for the initiation of the inflammation. On the other hand, *pyrin* controls to end the inflammation. The discovery of *ERAP* mutations in patients with FMF is not an unexpected finding. We believe that we showed the importance of having these two mutations at the same time to have clinically manifestation of FMF as a first time.

We believe that the evaluation of *ERAP* mutations in FMF patients with further studies will help us to find new tools for the correct diagnosis and current explanation the etiopathology of the disease. This study may help to establish a step to do this for further studies.

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All procedures performed in studies involving human participants were in accordance with the ethical standards of the institu-

tional and/or national research committee and with the 1964 Helsinki declaration and its later amendments or comparable ethical standards.

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