

procedures were used [14]. A xenograft is tissue transported between genetically dissimilar members of different species. It is osteo-conductive, biocompatible and structurally similar to human bone. There are two sources of xenografts used for bone replacement in periodontics: bovine bone and natural coral. A purported advantage of this product as a bone substitute is that it is natural in that it can provide structural components similar to that of human bone, improving its osteoconductive capability compared to that of synthetically derived mineral Aichelmann-Reidy and Yukna [15].

Bovine derived HA bone substitutes increase the available surface area that can act as an osteoconductive scaffold because of their porosity. This HA mineral content is comparable to that of bone, allowing it to become well vascularized and integrate with new host bone (Thaller, *et al.* 1994; Chen, *et al.* 1996) [16,17]. A statistically significant gain of clinical attachment and reduction in probing depth has been demonstrated when bovine bone was compared to a non-graft control for the treatment of human vertical osseous defects. When compared to demineralized freeze-dried bone allograft (DFDBA), similar amounts of probing depth reduction, clinical attachment level gain, bone fill and defect resolution were obtained Richardson [18].

Hence in this present study an effort was made to evaluate the effect of TI-OSS® bovine xenograft in treatment of intrabony defects with single flap approach in smokers and non-smokers. The present study showed significant reduction in the means of Plaque index (PI), sulcus bleeding index (SBI), probing depth (PD) and gain of Clinical attachment level (CAL) for both the smoker sites and

Non-smoker sites after 6 months, but the mean reduction of PI, CAL and SBI between the sites showed no statistical significant association after 6 months. This denotes the application of Bovine Xenograft did not reveal any additional benefit in the reduction of PI, CAL and SBI. These results are concomitant the study of Trombelli [19]. This should be because of the regular and frequent recall visits in which the patients underwent regular supra-gingival scaling and motivation for oral hygiene practice, which further minimized plaque accumulation. This implied that the participants of the study exercised good oral home care.

Periodontal pocket is deliberated as the pathognomonic sign of periodontal disease and reduction in probing pocket depth is one of the requisites for successful periodontal therapy. Probing depth

reduction in the present study was found to be significant from baseline to six months in the smokers' sites and Non-smoker sites.

Indicating that bovine xenograft has an added benefit on probing depth, this is similar to findings of a study of Yilmaz [20]. Ti-oss® is prepared from 100% cancellous bone without any cortical portion. Innovative pulverizing technique allows multi-porous structure, maximizing blood vessel ingrowth, Pre-Hydroxy Apatite structure, octacalcium phosphate crystal is found on the surface of Ti-oss®, resulting in fast bone formation. In the present study bone gain was 30.73% in smoker group sites and 53.64% in Non-smoker group sites ($p = 0.002$). This attributed the placement of bovine xenograft with single flap approach which provides a significant post-operative protection from exfoliation of the graft from the surgical site. The obtained results are in accordance with study conducted by Tombelli [19].

The purpose of developing these new techniques is to increase predictability, reduce patient discomfort, minimize the number of surgical sites and satisfy aesthetic demands of the treatment. When comparing early wound healing index at 2 weeks after the surgery, smokers showed a significantly lower number of sites with optimal wound healing and a higher number of sites with incomplete flap closure when compared to nonsmokers (0% vs 45.5% and 45.5% vs 18.2%, respectively), indicating a detrimental effect of smoking on early wound healing. This finding can be explained, at least in part by the alterations of the gingival vascular apparatus induced by the smoke [21,22] which in turn may have impaired blood perfusion. Blood perfusion was demonstrated to be a key determinant of the early healing of muco-periosteal flaps, and its impairment is frequently associated with wound dehiscence [23].

Treatment with bovine xenograft using single flap approach at defects resulted in substantial clinical attachment gain, with no significant intergroup difference. The magnitude of treatment effect is consistent with previous clinical trials investigating the use of bovine xenograft in the treatment of intraosseous defects [24]. In standardized periodontal defects experimentally created in rat molars, Azuma, *et al.* [25] demonstrated that the elevation of a single flap. A lower number of neutrophils, more rapid colonization of the elevated gingival tissues by fibroblasts, and greater connective tissue area occupied by type III collagen during early postoperative healing when compared to wide double flaps [26].

Another explanation for the lack of effect of smoking status can be related to the inclusive definition of smoker patient adopted in this study. A patient who smoked at least 1 cigarette per day was considered a smoker. The inclusion of light smokers may have mitigated, at least in part, the negative effect of smoking on the clinical outcomes.

This hypothesis seems to be corroborated, where patients smoking more than 10 cigarettes per day showed a clear tendency toward a lower CAL gain and PD reduction compared to patients smoking 1 to 10 cigarettes per day. In the light of our results, it was stated that bovine xenograft has yielded significant reduction in probing depth, gain in clinical attachment level (CAL) and significant radiographic bone fill, showing superior effects in treatment of intraosseous defects in smokers and non-smokers.

Graph 1: Comparison of mean difference of Plaque index (PI), Sulcus bleeding index (SBI), Probing depth (PD) and Clinical attachment level (CAL) between SMOKERS and NON-SMOKERS from baseline to 6 months.

Graph 2: Comparison of means of bone gain in smokers from baseline to 6 months.

Graph 3: Comparison of means of bone gain in non-smokers from baseline to 6 months.

Graph 4: Comparison of Means difference of Bone gain between Smokers and Non-Smokers from Baseline to 6 Months.

Variable	Time interval	Smokers	Non-Smokers
		Mean ± SD	Mean ± SD
Plaque index	Base line	2.13 ± 0.26	2.15 ± 0.26
	6 months	1.14 ± 0.22	1.09 ± 0.18
Sulcus bleeding Index	Base line	2.53 ± 0.52	2.53 ± 0.52
	6 months	1.47 ± 0.52	1.27 ± 0.46
Probing depth	Base line	8.4 ± 1.6	7.7 ± 1.2
	6 months	3.1 ± 0.5	3.6 ± 0.9
Clinical Attachment level	Base line	10.0 ± 1.9	10.1 ± 2.5
	6 months	5.5 ± 1.9	6.5 ± 2.0

Table 1: Means and Standard Deviations of Plaque Index (PI), Sulcus Bleeding Index (SBI), Probing Depth (PD) and Clinical Attachment Level (CAL) In Smokers and Non-Smokers at Different Time Intervals.

Variable	Time interval	Smokers Mean ± SD	Non-Smokers Mean ± SD
Plaque index	Base line to 6 months	1.06 ± 0.08 p = 0.001(S)	1.06 ± 0.08 p = 0.001(S)
Sulcus bleeding index	Base line to 6 months	1.26 ± 0.06 p = 0.000(S)	1.26 ± 0.06 p = 0.000(S)
Probing depth	Base line to 6 months	5.3 ± 1.3 p = 0.003(S)	4.1 ± 1.1 p = 0.003(S)
Clinical Attachment level	Base line to 6 months	4.5 ± 1.9 p = 0.003(S)	3.5 ± 0.09 p = 0.003(S)

Table 2: Comparison of Means of Plaque Index (PI), Sulcus Bleeding Index (SBI), Probing Depth (PD) and Clinical Attachment Level (CAL) in smokers and non-smokers within the sites from baseline to six months.

Statistical Analysis: Mann-Whitney U test.

S: Significant.

paired “t” test. Statistically significant if p < 0.05.

Variable	Smokers Mean ± SD	Non-Smokers Mean ± SD	p VALUE
Plaque index	1.06 ± 0.08	1.06 ± 0.08	0.467 NS
Sulcus bleeding index	1.26 ± 0.06	1.26 ± 0.06	0.148 NS
Probing depth	5.3 ± 1.3	4.1 ± 1.1	0.028 S
Clinical Attachment level	4.5 ± 1.9	3.5 ± 0.09	0.116 NS

Table 3: Comparison of mean difference of Plaque index (PI), Sulcus bleeding index (SBI), Probing depth (PD) and Clinical attachment level (CAL) between Smokers and Nonsmokers from baseline to 6 months.

Statistical Analysis: Independent ‘t’ test, paired t test.

Statistically significant if p < 0.05.

NS: Non-significant.

S: Significant.

Time interval	Smokers MEAN ± SD	Non-smokers MEAN ± SD	% of recovery	p VALUE
Base line	4.23 ± 0.94	4.53 ± 0.97	53.64	0.000S
Six months	1.30 ± 0.47	2.43 ± 0.57	30.73	0.000S

Table 4: Comparison of Means and standard deviation of Bone Gain in Smokers and Nonsmokers From Baseline to 6months.

Statistical analysis: paired “t” test. Statistically significant if p < 0.05.

Time interval	Smokers MEAN ± SD	Non-smokers MEAN ± SD	P value
Base line to 6 months	2.10 ± 0.40	2.93 ± 0.47	0.002 S

Table 5: Comparison of Mean Difference of Bone Gain in Smokers and Non-Smokers from Baseline to Six Months.

Statistical analysis: independent sample ‘t’ test. Statistically significant if p < 0.05.

S: Significant.

EHI score	Smokers	Non-Smokers
Complete flap closure, no fibrin line in the interproximal area	3	5
Complete flap closure, fine fibrin line in the interproximal area	3	4
Complete flap closure, fibrin clot in the interproximal area	5	3
Incomplete flap closure, partial necrosis of the interproximal tissue	1	0
Incomplete flap closure, complete necrosis of the interproximal tissue	0	0

Table 6: Early Healing Index (EHI) Assessment in smokers and non-smokers after 2 Weeks Following the Surgery.

Conclusion

We found that there was significant improvement in clinical parameters i.e., plaque index (PII), sulcus bleeding index (SBI),

probing depth (PD), clinical attachment level (CAL), and bone fill in the sites, before and after periodontal treatment. There was a significant improvement in clinical attachment level gain and bone fill in both the groups treated with bovine xenograft using a single flap approach from baseline to six months. The results of this study point towards the novel direction of current surgical regenerative approaches. Further, long term, multi-center, prospective longitudinal trials are the need of the hour to confirm the finding of this study.

Bibliography

1. Dieter D., et al. "Does periodontal tissue regeneration really work". *Periodontology 2000* 51 (2009): 208-219.
2. Yilmaz S., et al. "Treatment of intrabony periodontal defects with platelet rich plasma versus platelet poor plasma combined with a bovine derived xenograft: A controlled clinical trial". *Journal of Periodontology* 82 (2011): 837-844.
3. Giannobile WV and Somerman SJ. "Growth and amelogenin like factors in periodontal wound healing. A systematic review". *Annals of periodontology* 8 (2003): 193-204.
4. Owes HR., et al. "Platelet derived growth factor enhances demineralized bone matrix- induced cartilage and bone formation". *Calcified Tissue International* 42 (1988): 34-38.
5. Choukroun J., et al. "Platelet rich fibrin (PRF): A second generation platelet concentrate. Part IV: Clinical effects on tissue healing". *Oral Surgery, Oral Medicine, Oral Pathology, Oral Radiology, and Endodontology* 101 (2006): E56-60.
6. Centrella M., et al. "Transforming growth factor- β gene family members and bone". *Endocrine Review* 15 (1994): 27-39.
7. Gunja C., et al. "A boon for periodontal regeneration". *Journal of Advanced Research* 4 (2006): 396-401.
8. Takei HH., et al. "Flap technique for periodontal bone implants. papilla preservation technique". *Journal of Periodontology* 56 (1985): 204-210.
9. Cortellini P., et al. "The modified papilla preservation technique. A new surgical approach for interproximal regenerative procedures". *Journal of Periodontology* 66 (1995): 261-266.
10. Palmer RM., et al. "Mechanisms of action of environmental factors-tobacco smoking". *Journal of Clinical Periodontology* 32 (2005): 180-195.
11. Papanou PN and Tonetti M. "Diagnosis and epidemiology of periodontal lesions". *Periodontology 2000* 22 (2000): 8-21.
12. Trombelli L., et al. "Single flap approach with buccal access in periodontal reconstructive procedure". *Journal of Periodontology* 80 (2009): 353-360.
13. Farina R., et al. "Single flap approach in combination with enamel matrix derivative in the treatment of periodontal intraosseous defects". *International Journal of Periodontics and Restorative Dentistry* 34 (2014): 497-506.
14. Pradeep AR., et al. "Comparative evaluation of autologous platelet rich fibrin and platelet rich plasma in treatment of three walled intrabony defects in chronic periodontitis: A randomized controlled clinical trial". *Journal of Periodontology* 83 (2012): 1499-1507.
15. Aichelmann-Reidy ME and Yukna RA. "Bone replacement grafts, The bone substitutes". *Dental Clinics of North America* 42 (1998): 491-503.
16. Thaller SR., et al. "Repair of experimental calvarial defects with bio-oss particles and Collagen sponges in a rabbit model". *Journal of Craniofacial Surgery* 5 (1994): 242-246.
17. Chen L., et al. "Phenotypic catheterization of mononuclear cells following anorganic bovine bone implantation in rats". *Journal of Periodontology* 65 (1994): 1008-1015.
18. Richardson CR., et al. "Clinical evaluation of Bio-Oss: A bovine-derived xenograft for the treatment of periodontal osseous defects in humans". *Journal of Clinical Periodontology* 26 (1999): 421-428.
19. Trombelli L., et al. "Regenerative periodontal treatment with the single flap approach in smokers and smokers". *International Journal of Periodontics and Restorative Dentistry* 38 (2018): e59-e67.
20. Yilmaz S., et al. "Regenerative treatment with platelet-rich plasma combined with a bovine-derived xenograft in smokers

- and non-smokers:12-month clinical and radiographic results". *Journal of Clinical Periodontology* 37 (2010): 80-87.
21. Johnson GK and Guthmiller JM. "The impact of cigarette smoking on periodontal disease and treatment". *Periodontology* 2000 44 (2007): 178-194.
 22. Palmer RM., *et al.* "Mechanisms of action of environmental factors-Tobacco smoking". *Journal of Clinical Periodontology* 32 (2005): s180-s195.
 23. Zanetta-Barbosa D., *et al.* "Laser Doppler flowmetry of blood perfusion in mucoperiosteal flaps covering membranes in bone augmentation and implant procedures. A pilot study in dogs". *Clinical Oral Implants Research* 4 (1993): 35-38.
 24. Palachur D., *et al.* "A comparative evaluation of bovine-derived xenograft (bio-oss collagen) and type I collagen membrane (bio-gide) with bovine-derived xenograft (bio-oss Collagen) and fibrin fibronectin sealing system (tisseel) in the treatment of intrabony defects: A clinicoradio graphic study". *Journal of Indian Society of Periodontology* 18 (2014): 336-343.
 25. Azuma H., *et al.* "Single flap periodontal surgery induces early fibrous tissue generation by wound stabilization". *Journal of Hard Tissue Biology* 26 (2017): 119-126.
 26. Wennström JL and Lindhe J. "Some effects of enamel matrix proteins on wound healing in the dento-gingival region". *Journal of Clinical Periodontology* 29 (2002): 9-14.

Assets from publication with us

- Prompt Acknowledgement after receiving the article
- Thorough Double blinded peer review
- Rapid Publication
- Issue of Publication Certificate
- High visibility of your Published work

Website: www.actascientific.com/

Submit Article: www.actascientific.com/submission.php

Email us: editor@actascientific.com

Contact us: +91 9182824667