# ACTA SCIENTIFIC AGRICULTURE (ISSN: 2581-365X)

Volume 2 Issue 12 December 2018

# Ameliorative Effects of Exogenously Applied Nitric Oxide on Seed Germination and Seedling Growth of Purslane Under Drought and Salinity

# Melek Ekinci<sup>1</sup>, Selda Ors<sup>2</sup>, Metin Turan<sup>2</sup>, Raziye Kul<sup>1</sup> and Ertan Yildirim<sup>1\*</sup>

<sup>1</sup>Atatürk University, Agriculture Faculty, Department of Horticulture, Erzurum, Turkey <sup>2</sup>Atatürk University, Faculty of Agriculture, Department of Agricultural Structures and Irrigation, Erzurum, Turkey **\*Corresponding Author:** Ertan Yildirim, Atatürk University, Agriculture Faculty, Department of Horticulture, Erzurum, Turkey.

Received: October 30, 2018; Published: November 06, 2018

## Abstract

We investigated the effects of exogenously applied nitric oxide (NO) on seed germination and seedling properties of purslane (*Portulaca oleracea* L.) under different levels of drought and salinity. The seeds were soaked in different SNP (sodium nitroprusside as a NO donor) doses (0, 50, 100, 150 and  $200 \,\mu$ M) during 12 h. Drought and salinity stress were generated by additions of PEG 6000 or sodium chloride to medium. Water potentials were: 0, -0.20, -0.75, and -1.53 MPa for both stresses. The seeds were allowed to germinate at 25oC, germination percentage, germination speed, main daily germination, root and shoot length were determined at the end of the experiment. Germination and seedling properties were inversely proportional to the concentrations of NaCl and PEG, thus purslane showed a reduction in germination and seedling growth with increased NaCl or PEG concentrations, but the reduction in PEG treatment were higher than NaCl treatment. However, exogenously NO treatments alleviated the negative impact of stress conditions for germination and seedling characteristics. 50, 100 and 200  $\mu$ M doses of NO were the most efficient treatments on ameliorating the deleterious effects of salinity and drought. It can be concluded that NO applications may reduce the negative effects of moderate and high salt and drought stress during germination and early growth stage.

Keywords: Abiotic Stress; Water Deficit; Salinity; Nitric Oxide; Germination; Portulaca Oleracea

## Introduction

Abiotic stress factors affect plant growth significantly and cause significant decreases in productivity and quality in agricultural produce. Environmental stress factors such as drought, salinity, low and high temperatures are important abiotic stresses which limit agricultural production [1]. The most important stress factors that cause damage to agricultural production are known as drought and salinity in the world [1,2].

High salt concentration causes osmotic and ion stress in plants and increase the leakage of electrons from the transport of electrons in chloroplasts and mitochondria and causes excessive production of reactive oxygen species (ROS) in plants [3-5]. Salinity reduces the growth rate resulting in smaller leaves, shorter lengths and sometimes fewer leaves [1]. Drought conditions have a negative effect on photosynthetic activity, cause a change in chlorophyll content and components in the cell, decrease plant root and shoot growth, inhibit photochemical activities and decrease the activity of enzymes in Calvin circle [6-10].

NO, an important signal molecule, is a product of nitrogen metabolism known as a stress regulator in signal transduction pathways [11]. NO is stated to have multiple biological roles in developmental processes such as germination, root organogenesis, stoma closure, flowering and senescence [12]. In addition, NO was found to be involved in environmental responses such as drought, temperature, salinity, heavy metal and UV-B radiation in plants [2,13-18].

NO decreases the oxidative damage caused by salinity in seedlings and increases the growth in the seedlings and dry weight under salt and drought stress [2,19,20]. In addition, the researchers reported that the pretreatment of NO in plants under stress condi-

tions increases the total soluble protein, effectively contributing to a better balance between carbon and nitrogen metabolism [20,21]. NO affects the downstream of ABA in the adaptation reaction to water-deficiency stress by inducing stoma closure [22]. NO increases the capacity of antioxidants, stability of cellular membranes and improves the photosynthetic capacity [23].

Purslane is providing both novel biologically active substances and essential compounds for human nutrition [24]. The purslane plant was found to be 6.3 dS m<sup>-1</sup> (EC) threshold and moderately tolerant to salinity [25]. Also, it was determined that purslane is a drought-tolerant plant [24]. Purslane outstanding tolerance to chloride salinity makes this species a promising halophyte candidate for saline agriculture [26].

The current scientific literature demonstrates that NO usage is one-way to improve the plant's tolerance on stress conditions. However, the effect of NO on seed germination and seedling growth of purslane under drought and salinity stress conditions has yet not been well studied. Thus, this experiment focuses on the effect of the exogenous seed application of NO on seed germination and seedling growth properties of purslane under saline and growth conditions.

#### **Material and Methods**

Purslane (*Portulaca oleracea* L.) was used as a plant material. One percent sodium hypochlorite was used for seed disinfection about two minutes, and then seeds were washed with distilled water immediately. Seeds were soaked in different SNP (sodium nitroprusside) solutions (0, 50, 100, 150 and 200  $\mu$ M) for 12 hours in a germinator with a temperature of 23 ± 2°C. Treated seeds were dried on the papers under room temperature. Fifty disinfected seeds treated with NO were germinated in 2 folds of Whatman No. 1 filter paper (sterilized) which were placed in Petri dishes (15 cm diameter). Each dish was moistened with 10 ml solutions containing polyethylene glycol (PEG) 6000 or sodium chloride (NaCl) solutions with osmotic potential 0, -0.20, -0.75 and -1.53 MPa. In the control group only, distilled water was used.

The germination study was carried out with 4 replications and 50 seeds were used in each replication [27]. Seeds were allowed to germinate at 25°C. Germination percentage (%) and germination speed [28], mean daily germination [29], root length (mm) and shoot length (mm) were measured at the end of the study (14 day). The germination percentage values were transformed to arc sin before ANOVA. The data were analyzed by means of Duncan multiple comparison test [30].

#### Results

Effects of NO treatments on germination speed, germination percentage and daily mean germination of purslane seeds under drought and salinity conditions are given in table 1. The effect of salinity, drought and NO applications on seed germination speed was found to be statistically significant (p < 0,001). With increasing salinity and drought severity, the germination speed values increased, and germination occurred later in the highest stress conditions than the control. According to the averages, the latest germination was in the 0 µM NO application, while the earliest germination occurred in 200 µM NO application in salinity conditions. On the other hand, in drought conditions the latest germination was in the 0 µM NO application, while the earliest germination occurred in 100  $\mu$ M NO (Table 1). The 50 and 100  $\mu$ M NO applications resulted in faster germination at -0.20 MPa and -0.75 MPa salt stresses, respectively. In drought conditions, the 150  $\mu$ M NO applications provided the earliest germination at -0,20 MPa and -0,75 MPa stress, while 100 µM NO application showed faster germination at -1,53 MPa drought stress (Table 1).

17

The effect of salinity, drought and NO applications on seed germination percentage was found to be statistically significant (p < 0,001). With increasing salinity and drought, the germination percentage decreased in the highest stress conditions. According to the averages, the highest germination percentage was in the 150 µM NO application, while the lowest germination percentage occurred in 0 µM NO application in salinity stress conditions. On the other hand, in drought conditions the highest germination percentage was in the 100 µM NO application, while the lowest germination occurred in 200 µM NO (Table 1). The highest germination percentage was obtained from 150 µM NO application at -0.20 MPa and -0.75 MPa salt stresses and 50  $\mu$ M NO application at -1,53 MPa salt stress condition. On the other hand, the highest germination percentage was obtained from 100 µM NO application at -0.20 MPa, 50 µM NO application at -0.75 MPa salt stresses and 50 µM NO and 100 µM NO applications at -1,53 MPa drought stress condition (Table 1).

The effects of applications on mean daily germination of purslane seed were shown in table 1. Increased levels of salt and drought reduced mean daily germination. The highest mean daily germination was obtained 150  $\mu$ M NO application for salinity and 100  $\mu$ M NO application for drought conditions (Table 1). In this study, the highest mean daily germination occurred in150  $\mu$ M NO application at -0.20 MPa and -0.75 MPa salt stress levels, while the highest mean daily germination was obtained from 100  $\mu$ M NO ap-

			Germinatio	n Speed			
Stress	Doses	0-NO	50-NO	100-NO	150-NO	200-NO	Mean
Salt	0	1,37 d	1,31 d	1,29 d	1,18 d	1,32 d	1,29 C
	-0,20 MPa	1,35 d	1,15 d	1,30 d	1,19 d	1,17 d	1,23 C
	-0,75 MPa	1,86 c	1,78 c	1,69 c	1,84 c	1,74 c	1,78 B
	-1,53 MPa	3,81 b	3,86 b	4,29 a	3,89 b	3,82 b	3,93 A
Mean		2,10 <sup>ns</sup>	2,02	2,14	2,03	2,01	
Application	n x Salt	P < 0,001					
	0	2,27 d	1,80 def	1,72 def	1,99 def	1,81 def	1,92 C
	-0,20 MPa	1,83 def	1,66 ef	1,56 f	1,52 f	1,64 ef	1,64 D
Drought	-0,75 MPa	2,26 d	2,17 de	2,26 d	2,05 def	2,17 de	2,18 B
	-1,53 MPa	4,83 a	4,68 a	3,25 c	3,94 b	4,00 b	4,14 A
Mean		2,80 A	2,58 AB	2,20 C	2,37 BC	2,41 BC	
Application	n x Drought	P < 0,001					
		Ger	mination Per	centage (%)			
Salt	0	94,00 bcd	96,00 ab	98,00 a	97,00 ab	95,00 bc	96,00 A
	-0,20 MPa	93,50 bcd	93,50 bcd	95,50 abc	96,00 ab	94,50 bc	94,60 B
	-0,75 MPa	90,50 e	91,00 de	91,50 cde	94,50 bc	91,00 de	91,70 C
	-1,53 MPa	77,00 f	78,00 f	77,00 f	77,00 f	75,00 f	76,80 D
Mean		88,75 C	89,63 BC	90,50 B	91,13 A	88,88 BC	
Application	n x Salt	P < 0,05					
	0	94,50 cd	96,00 bc	98,50 a	95,00 bc	98,00 ab	96,40A
Drought	-0,20 MPa	94,00 cd	95,50 bc	97,50 ab	95,00 bc	94,00 cd	95,20 A
	-0,75 MPa	93,00 cd	95,50 bc	92,00 cd	95,00 bc	90,00 d	93,10 B
	-1,53 MPa	9,00 ef	12,00 e	12,00 e	8,00 ef	6,00 f	9,40 C
Mean		72,63 C	74,75 AB	75,00 A	73,25 BC	72,00 C	
Application	n x Drought	P < 0,05					
			Mean Dail	y Germinatio	n		
Salt	0	6,71 a-d	6,86 ab	6,86 ab	6,93 a	6,79 ab	6,83 A
	-0,20 MPa	6,68 b-e	6,64 b-e	6,82 ab	6,86 ab	6,75 abc	6,75 A
	-0,75 MPa	6,46 ef	6,50 def	6,54 c-f	6,68 b-e	6,43 f	6,52 B
	-1,53 MPa	5,50 g	5,43 g	5,50 g	5,50 g	5,36 g	5,46 C
Mean		6,34 B	6,36 B	6,43 AB	6,49 A	6,33 B	
Application x Salt		P < 0,05					
Drought	0	6,64 cd	6,82 abc	7,04 a	6,79 abc	6,43 d	6,74 A
	-0,20 MPa	6,71 bc	6,82 abc	6,96 ab	6,78 abc	6,71 bc	6,80 A
	-0,75 MPa	6,64 cd	6,82 abc	6,57 cd	6,78 abc	7,00 a	6,76 A
	-1,53 MPa	0,61 ef	0,86 e	0,86 e	0,61 ef	0,43 f	0,67 B
Mean		5,15 B	5,33 A	5,36 A	5,24 AB	5,14 C	
Application	n x Drought	P < 0,001					

**Table 1:** The effects of NO applications on germination properties of purslane under salinity and drought conditions.

ns: non-significant, different lower case letters indicate differences related to NO and stress applications, and different capital letters indicate differences related to means.

**Citation**: Ertan Yildirim., et al. "Ameliorative Effects of Exogenously Applied Nitric Oxide on Seed Germination and Seedling Growth of Purslane Under Drought and Salinity". Acta Scientific Agriculture 2.12 (2018): 16-22.

18

19

plication at -0.20 MPa, with 200  $\mu$ M NO application at -0.75 MPa drought stress levels (Table 1).

and salt stress have caused a decrease in seedling root and shoot length. The root and shoot lengths could not measure at the highest level of drought stress factor (-1.53 MPa level). The effect was found statistically important (p < 0,001) for both stress factors.

The effects of applications on seedling root and shoot length under stress conditions were given in table 2. The severe drought

			Root Leng	gth (mm)			
Stress	Doses	0-NO	50-NO	100-NO	150-NO	200-NO	Mean
Salt	0	2,95 b	3,14 a	2,95 b	3,03 ab	2,70 c	2,95 A
	-0,20 MPa	2,36 de	2,67 c	2,24 e	2,50 d	1,93 f	2,34 B
	-0,75 MPa	1,24 h	1,91 f	1,30 h	1,60 g	1,57 g	1,52 C
	-1,53 MPa	0,22 1	0,28 1	0,14 1	0,13 ı	0,181	0,19 D
Mean		1,69 C	2,00 A	1,66 CD	1,81 B	1,59 D	
Application x Salt		P < 0,001					
Drought	0	3,68 abc	3,96 ab	3,48 bcd	3,42 cd	4,10 a	3,73 A
	-0,20 MPa	3,63 abc	3,94 ab	3,27 cd	3,21 cd	4,01 a	3,61 A
	-0,75 MPa	2,22 g	2,71 ef	2,34 fg	2,51 fg	3,08 de	2,57 B
	-1,53 MPa	-	-	-	-	-	-
Mean		3,18 B	3,53 A	3,03 B	3,05 B	3,73 A	
Application x Drought		P < 0,05					
			Shoot Len	gth (mm)	1		
Salt	0	2,64 ab	2,72 a	2,71 ab	2,68 ab	2,55 bc	2,66 A
	-0,20 MPa	2,61 ab	2,65 ab	2,45 c	2,56 abc	2,44 c	2,54 B
	-0,75 MPa	2,00 de	1,92 e	1,86 e	1,99 e	2,15 d	1,98 C
	-1,53 MPa	0,69 f	0,78 f	0,62 f	0,68 f	0,64 f	0,68 D
Mean		1,99 AB	2,02 A	1,91 AB	1,97 AB	1,94 AB	
Application x Salt		P < 0,01					
Drought	0	2,26 b	2,42 ab	2,55 a	2,64 a	2,42 ab	2,46 A
	-0,20 MPa	2,03 c	1,67 ef	1,90 cd	1,99 c	1,85 cde	1,89 B
	-0,75 MPa	1,23 h	1,63 efg	1,41 gh	1,56 fg	1,70 def	1,51 C
	-1,53 MPa	-	-	-	-	-	
Mean		1,84 C	1,91 BC	1,95 ABC	2,06 A	1,99 AB	
Application x Drought		P < 0,001					

Table 2. The effects of NO applications on root and shoot length of purslane on salinity and drought conditions.

Different lower case letters indicate differences related to NO and stress applications and different capital letters indicate differences related to means.

NO applications had important impact on seedling root and shoot length. For salinity condition, the highest root length was obtained from 50  $\mu$ M NO application for all stress levels. On the other hand, for drought condition the highest root length was occurred in 200  $\mu$ M NO for -0.20 MPa and -0,75 MPa levels. 50  $\mu$ M NO application showed the highest shoot length at -0.20 MPa and -1.53 MPa salinity levels, while 200  $\mu$ M NO application gave the highest shoot length at -0,75 MPa drought levels.

#### Discussion

Salinity and drought stress conditions hindered the investigated parameters in this experiment. Germination properties of purslane seeds were adversely affected by salinity and drought stress. Salinity and drought stresses are physiologically related, because both induce osmotic stress and most of the metabolic responses of the affected plants are similar to some extent. Both

stress conditions may cause significant reductions in the rate and final percentage of germination [31]. Similarly, drought and salinity stress conditions negatively affected the shoot and root length. At zero potential, both shoot and root lengths reached their highest values. All other treatments gradually reduced the seedling growing. There are a lot of studies showing that stress conditions lead to decrease in growth properties of crops [7,9]. The first measurable effect resulting from drought and salt stress is the decline in growth. This is due to the cessation of cellular expansion. Cellular expansion process has been reported to be susceptible to salinity and drought stress conditions [32].

To investigate the interactive role of NO on the drought and salinity, we applied 50, 100, 150 and 200  $\mu$ M NO, and found that NO treatments mitigated the adverse effect of stress conditions on germination and seedling growth. As a result of the study, the effect of NO applications in the purslane germination attributes is not much in stress-free conditions. Exogenous NO has a strong stimulating effect on seed germination under stress or non-stress conditions [33]. Similarly, it was observed NO treatments on seed alleviated stress conditions on germination properties in different crops by enhancing antioxidant capability [3,34]. Exogenous NO application has effective treatment to protect plants from oxidative damage under salt and drought stress by increasing various antioxidant defense enzymes activity (SOD, APX, CAT and GR) [3,35].

The purslane seeds are relatively salinity and drought tolerant, but especially -1.53 MPa severity of salinity and drought caused significant reductions on germination and root and shoot length of seed. However, 50 µM doses of NO at the most severe salt level and 50 and 100  $\mu$ M doses of NO at the most severe drought level mitigated the negative impact of stress. In terms of root and shoot lengths, the negative effect of stress was less in 50 µM NO application at the highest salinity and in 50 µM and 200 µM NO applications at the highest level of drought as compared to the other applications. This situation can be explained by the fact that NO has antioxidant properties and can act as a signal in activating ROSscavenging enzyme activities under stress [20]. In addition to the regulatory role in some physiological processes such as germination, flowering, fruit ripening and senescence, NO plays a protective role against different abiotic stresses such as drought, salinity, temperature [36,37].

In the study, it was found out that drought is more deleterious than salinity on observed parameters of purslane. Root and shoot measurement could not be performed in level of -1.52 MPa of drought. This also shows that purslane is more resistant to salinity than drought during germination and seedling. Kumamoto., et al. [25] reported that the purslane was moderately tolerant to salinity. And also, the researchers suggest that purslane could be used to rehabilitate saline soils and halophyte candidate for saline agriculture [26,38,39].

20

#### Conclusion

As a result of the study, germination properties of purslane under salinity and drought conditions slightly decreased at lower levels of stress, but drastically decreased at the highest stress levels. However, the seedling growth decreased gradually depending on increased stress conditions. It can be concluded that purslane is relatively resistant to low and moderate salt and drought stress during germination and early seedling growth. NO applications can mitigate the negative effects of moderate and high salt and drought stress.

## **Bibliography**

- 1. Meena MD., *et al.* "Effects of municipal solid waste, rice straw composts and mineral fertilizers on biological and chemical properties of a saline soil and yields of mustard pearl millet cropping system". *Soil Research* 54.8 (2016): 958-966.
- Garcia-Mata C and Lamattina L. "Nitric Oxide Induces Stomatal Closure and Enhances the Adaptive Plant Responses against Drought Stress". *Plant Physiology* 126 (2001): 1196-1204.
- Zheng C., *et al.* "Exogenous nitric oxide improves seed germination in wheat against mitochondrial oxidative damage induced by high salinity". *Environmental and Experimental Botany* 67 (2009): 222-227.
- 4. Haihua R., *et al.* "Protective effects of nitric oxide on salt stressinduced oxidative damage to wheat (Triticum *aestivum* L.) leaves". *Chinese Science Bulletin* 47.8 (2002): 677-681.
- Sheokand S., *et al.* "Effect of nitric oxide and putrescine on antioxidative responses under NaCl stress in chickpea plants". *Physiology and Molecular Biology of Plants* 14.4 (2008): 355-362.
- 6. Monakhova OF and Chernyadev II. "Protective role of kartolin-4 in wheat plants exposed to soil drought". *Applied and Environmental Microbiology* 38 (2002): 373–380.
- Gallardo M., *et al.* "Response of shoot diameter to water stress in greenhouse-grown vegetable crops". *Acta Horticulture* 664 (2004): 253-260.

- 8. Anjum SA., *et al.* "Morphological, physiological and biochemical responses of plants to drought stress". *African Journal of Agricultural Research* 6 (2011): 2026-2032.
- 9. Osakabe Y., *et al.* "Response of plants to water stress". *Front Plant Science* 5 (2014): 86.
- **10.** Razzaghi F., *et al.* "Ionic and photosynthetic homeostasis in quinoa challenged by salinity and drought mechanisms of tolerance". *Functional Plant Biology* **42**.2 (2014): 136-148.
- 11. Besson-Bard A., *et al.* "New insights into nitric oxide signaling in plants". *Annual Review of Plant Biology* 59 (2008): 21-39.
- 12. Lamattina L., *et al.* "Nitric oxide: the versatility of an extensive signal molecule". *Annual Review of Plant Biology* 54 (2003): 109-136.
- 13. Arasimowicz-Jelonek M., *et al.* "Involvement of nitric oxide in water stress-induced responses of cucumber roots". *Plant Science* 177 (2009): 682-690.
- 14. Song LL., *et al.* "Nitric oxide protects against oxidative stress under heat stress in the calluses from two ecotypes of reed". *Plant Science* 171 (2006): 449-458.
- 15. Zhao MG., *et al.* "Nitric oxide synthase-dependent nitric oxide production is associated with salt tolerance in *Arabidopsis*". *Plant Physiology* 144 (2007): 206-217.
- 16. Rodríguez-Serrano M., *et al.* "Cadmium effect on oxidative metabolism of pea (*Pisum sativum* L.) roots. Imaging of reactive oxygen species and nitric oxide accumulation *in vivo*". *Plant Cell Environment* 29 (2006): 1532-1544.
- 17. Ma W., *et al.* "Nitric oxide modulates cadmium influx during cadmium-induced programmed cell death in tobacco BY-2 cells". *Planta* 232 (2010): 325-335.
- Shi S., *et al.* "Protective effect of nitric oxide against oxidative stress under ultraviolet-B radiation". *Nitric Oxide* 13 (2005): 1-9.
- 19. Song J., *et al.* "Effects of nitric oxide and nitrogen on seedling emergence, ion accumulation, and seedling growth under salinity in the euhalophyte Suaeda salsa". *Journal of Plant Nutrition and Soil Science* 172 (2009): 544-549.
- 20. Siddiqui MH., *et al.* "Role of nitric oxide in tolerance of plants to abiotic stress". *Protoplasma* 248 (2011): 447-455.
- 21. Zheng C., *et al.* "Effects nitroprusside, a nitric oxide donor, on carbon and nitrogen metabolism and the activity of the anti-oxidation system in wheat seedlings under salt stress". *Acta Ecologica Sinica* 30 (2010): 1174-1183.

22. Planchet E., *et al.* "Abscisic acid-induced nitric oxide and proline accumulation in independent pathways under water-deficit stress during seedling establishment in *Medicago truncatula*". *Journal of Experimental Botany* 65.8 (2014): 2161-2170.

21

- 23. Rahimian Boogar A., *et al.* "Exogenous nitric oxide alleviates oxidative damage in turfgrasses under drought stress". *South African Journal of Botany* 92 (2014): 78-82.
- 24. Anastacio A and Carvalho S. "Accumulation of fatty acids in purslane grown in hydroponic salt stress conditions". *International Journal of Food Sciences and Nutrition* 64.2 (2013): 235-242.
- 25. Kumamoto J., *et al.* "Purslane: a potential new vegetable crop rich in omega-3 fatty acid with controllable sodium chloride content. In: Naqvi, H.H., Estilai, A., Ting, I.P (Eds). First International Conference on New Industrial Crops and Products. University of Arizona, Riverside CA, Tucson, AZ, (1990): 229-233.
- 26. Kafi M and Rahimi Z. "Effect of salinity and silicon on root characteristics, growth, water status, proline content and ion accumulation of purslane (*Portulaca oleracea* L.)". Soil Science and Plant Nutrition 57 (2011): 341-347.
- 27. ISTA. "International rules for seed testing". *Seed Science Technology* 21 (1999): 288.
- Ellis RH and Roberts EH. "The quantification of ageing and survival in orthodox seeds". *Seed Science Technology* 9 (1981): 373-409.
- 29. Czabator FJ. "Germination value: An index combining speed and completeness of pine seed germination". *Forest Science* 8 (1962): 386-395.
- 30. SPSS Inc. SPSS® 18.0 Base User's Guide, Prentice Hall (2010).
- 31. Farsiani A and Ghobadi ME. "Effects of PEG and NaCl stress on two cultivars of corn (*Zea mays* L.) at germination and early seedling stages". *World Academy of Science, Engineering and Technology* 57 (2009): 382-385.
- 32. Shalhevet J., *et al.* "Root and shoot growth responses to salinity in maize and soybean". *Agronomy Journal* 87 (1995): 512-516.
- 33. Zhang H. "Nitric Oxide Alleviates the Inhibition of Salinity Stress on Seed Germination and Seedling Growth of Cynanchum bungei Decne (Asclepiadaceae)". *HortScience* 50 (2015):119-122.

22

- Fan HF., *et al.* "Effects of nitric oxide on the germination of cucumber seeds and antioxidant enzymes under salinity stress". *Acta Physiology Plant* 35 (2013): 2707-2719.
- 35. Shehab GG., *et al.* "Effects of various chemical agents for alleviation of drought stress in rice plants (*Oryza sativa* L.)". *Notulae Botanicae Horti Agrobotanici Cluj-Napoca* 38 (2010): 139-148.
- Arasimowicz M., *et al.* "Nitric oxide as a bioactive signalling molecule in plant stress responses". *Plant Science* 172 (2007): 876-887.
- Habib N., *et al.* "Enhancement in seed germinability of rice (Oryza *sativa* L.) by pre-sowing seed treatment with nitric oxide (NO) under salt stress". *Pakistan Journal of Botany* 42 (2010): 4071-4078.
- Hamidov A., et al. "Environmentally useful technique-Portulaca oleracea golden purslane as a salt removal species". WSEAS Transactions on Environment and Development 3.7 (2007): 117-122.
- Meena., *et al.* "Changes of phosphorus fractions in saline soil amended with municipal solid waste compost and mineral fertilizers in a mustard-pearl millet cropping system". *Catena* 160 (2018): 32-40.

# Volume 2 Issue 12 December 2018 © All rights are reserved by Ertan Yildirim., *et al.*